

**AKENTEN APPIAH- MENKA UNIVERSITY OF SKILLS TRAINING AND  
ENTREPRENEURIAL DEVELOPMENT**

**EFFECT OF PARTIAL REPLACEMENT OF MAIZE WITH FULL-FAT SOYA  
BEAN OR SOYA BEAN OIL ON THE GROWTH PERFORMANCE, GUT pH,  
CARCASS AND BONE TRAITS OF ROSS 308 BROILER CHICKENS**

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**2025**

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ENTREPRENEURIAL DEVELOPMENT**

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of the requirements for the award of the degree of Master of Philosophy Animal  
Science (Animal Production and Management)**

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# DECLARATION

## Student's Declaration

I, Ampoma Samuel, declare that this thesis, with the exception of quotations and references contained in published works which have all been identified and duly acknowledged, is entirely my own original work, and it has not been submitted, either in part or whole, for another degree elsewhere.

Ampoma Samuel (Student)

Signature.....

Date: ...../...../.....

## Supervisor's Declaration

I hereby declare that the preparation and presentation of this work was supervised in accordance with the guidelines for supervision of thesis as laid down by the Akenten Appiah- Menka University of Skills Training and Entrepreneurial Development.

Associate Prof. Holy Kwabla Zanu

Signature.....

Date: ...../...../.....

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## **DEDICATION**

I dedicate this thesis to my late brother Obeng Asamoah Benjamin and my lovely wife, children and all my family members.

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## **LIST OF ABBREVIATION**

<b>ABBREVIATION</b>	<b>DEFINITION</b>
ADFI	Average daily feed intake
ANFs	Anti-nutritional factors
BDM	Bone mineral density
BW	Body weight
CP	Crude protein
DM	Dry matter
EU	European Union
FCR	Feed conversion ratio
FFSB	Full fat soya bean
ME	Metabolisable energy
NSP	Non-starch polysaccharides
PA	Phytic acid
SBM	Soya bean meal
SCFA	Short chain fatty acid

## ABSTRACT

Soya bean (SB) products are valuable protein and energy source commonly used in poultry diets. Thus, this study was designed to investigate the effect of partial replacement of maize with full-fat soya beans or soya bean oil on growth performance, gut pH, carcass traits and bone characteristics of Ross 308 broiler chickens. A total of 200 broiler chickens were randomly assigned to four dietary treatments T1(regular soya bean diet), T2(full fat soya bean diet), T3(soya bean oil diet) and T4 (regular soya bean diet plus full fat soya bean diet) in a completely randomized design (CRD), with five replicate, ten birds per pen and fifty birds per treatment. Growth performance indicators such as body weight, body weight gain, feed intake, livability, and feed conversion ratio were assessed every two weeks. Gut pH, carcass traits, bone characteristics were also measured. Results showed that on d 14 there was no significant ( $P > 0.05$ ) effect of the treatment on all the growth parameters measured, numerically, all the growth parameters increased but on d 28 of the study, dietary treatment had a significant ( $P < 0.05$ ) influence on feed intake which increased with birds on T3 recording the highest feed intake but statistically was similar to T4 and T1 ( $P < 0.05$ ). Also, from d 42 to 56 there was an increase with a significant difference on dietary treatments on body weight, body weight gain and feed intake of the birds ( $P < 0.05$ ). Dietary treatments had no significant effect ( $P > 0.05$ ) on gut pH across all segments. Gizzard and fat pad were significantly ( $P < 0.05$ ) influenced by the dietary treatment. Regarding bone traits, significant differences ( $P < 0.05$ ) were observed only in tibia length, with birds on T1, T2, and T4 showing longer tibiae than those on T3. In terms of cost benefits, there was significant difference ( $P < 0.05$ ) between the treatment means. Birds on soya bean oil diet (T3) and conventional plus full-fat soya bean diets (T4) resulted in

increased ( $p < 0.05$ ) price per bird, profit, and Performance Efficiency Index (PEI) compared to the birds fed full-fat soya bean diet. In conclusion, using soya bean oil or full-fat soya bean as an alternative energy source by reducing the maize in a diet had positive impact on the growth performance especially at the early stage, carcass traits and tibia length of the birds were optimal.

# CHAPTER ONE: INTRODUCTION

## 1.1 Background to the Study

Globally, maize and soya bean are primarily used as the main ingredients in chicken feed. Maize grain remains the main source of energy in commercial broiler chicken diets but the price of maize has increased significantly over time due to intense competition for its usage by man or other livestock species and industries (Erenstein, *et al.*, 2022). Maize constitutes about 60-70 % in most poultry diet and has a greater acceptance in poultry feeds (Dei *et al.*, 2017). Thus, it is imperative to search for suitable alternative sources of energy in poultry feeds to reduce the cost of production.

Several ingredients like cassava, sorghum, wheat bran, full-fat soya bean, soya bean oil have been identified as suitable ingredients to partially or completely replace maize in poultry diet in an attempt to cut down feed cost (Toomer, 2024). Most of these ingredients that could replace maize contain absorption anti nutritional factors (ANFs) which might hinder the digestion and of nutrients in chicken (Ababor *et al.*, 2023). Elsewhere, vegetable oils are added to chicken feed as a source of energy to reduce the quantity of maize in an attempt to reduce the cost of feeding but in Ghana vegetable oil is not used in formulating chicken diet (Ashong *et al.*, 2024).

Soya bean is one of the locally produced protein sources for chicken in Ghana. The oil extracted from soya bean is used for domestic consumption and export (Hailu, *et al.*, 2014). However, through diet manipulation, full-fat soya bean or soya bean oil can be used to partially replace maize in commercial chicken diets (Toomer *et al.*, 2024). Soya bean oil is produced primarily for human consumption. However, it has become a useful source of

feed-grade fat for animals due to a need to formulate high-energy diets for modern breeds. Feed-grade soya bean oil is popularly used in high energy diets, particularly for poultry, because of its high digestibility and metabolizable energy content compared with other vegetable fats/oils (Bergeron, 2021). It is used widely in rations for broiler chickens and growing turkeys as a feed-grade fat to increase energy density of feeds and improve efficiency of feed utilization (Babatunde *et al.*, 2021). The high energy value of soya bean oil is attributed to its high percentage of (poly) unsaturated fatty acids, which are well absorbed and utilized as a source of energy by the animal (Castro *et al.*, 2019).

## **1.2 Problem Statement**

In recent years, cost of producing broiler birds has gone up due to high cost of ingredient such as maize. There is always shortage of maize because of competition on the usage of maize by man, climate change and other production challenges. Dei (2017) The search for cheaper sources of energy will reduce cost of production of broilers. Ahiwe *et al* (2018). Poultry feed formulation is customized to suit the specific genetic strain, age, and environmental factors influencing the birds. Typically, it comprises a balanced blend of grains, protein supplements, vitamins, minerals, and additives to meet the birds' nutritional requirements Dei. (2017) This has compelled researchers to delve into less cost non - conventional feed sources such as full fat soya bean or soy oil (FFS or SO). Therefore, the purpose of this study was to assess the effects of partial replacement of maize with full fat soya bean or soy oil on growth performance, gut pH, bone breaking strength, and carcass traits of Ross 308 broiler chickens.

### **1.3 Main Objective**

The main objective of the study was to investigate soya bean oil or full-fat soya bean as alternative source of energy to maize in broiler diets.

#### **1.3.1 Specific objectives:**

Specifically, the study sought to evaluate the effects of the treatment on:

- i. Growth performance
- ii. Bone strength
- iii. Gut pH
- iv. Carcass traits
- v. Profitability margin

## CHAPTER TWO: LITERATURE REVIEW

### 2.1 Maize production in Ghana

Maize is one of the most versatile and emerging cereal crops having wider adaptability under varied agro-climatic conditions (Kumar and Jhariya, 2013). There are about 50 species, with a variety of colours, textures, and sizes and forms of grain. Red, yellow, and white are the most widely grown varieties of maize. Animal feed has made significant use of the white and yellow types, which are the most popular (Prasanna, 2012). Maize belongs to the family Poaceae and is widely consumed as a staple food in many countries around the world (Shah *et al.*, 2016). In Ghana, it is considered the second most important staple food next to cassava (Murdia *et al.*, 2016). Maize is the main input in feed production for poultry. Andam *et al.* (2017) reported that over the past five years, Ghana's annual production averaged 1.8 million metric tons per year harvested from approximately 1.02 million hectares. Globally, maize is known as queen of cereals because it has the highest genetic yield potential among the cereals (Mwambo *et al.*, 2020).

According Dei (2017), maize constitutes about 50-60% in most poultry diets and has a greater acceptance in poultry feeds. But its production is not sufficient to meet the ever-increasing demand of poultry industry. Also, its price is increasing continuously due to intense competition for its usage by man or other livestock species (Erenstein *et al.*, 2022) and starch and allied industries. Maize is a conventional energy source and is currently, the most widely used grain crop in the Ghanaian poultry industry because it provides the bulk of most poultry diets. In terms of total cost, energy is the most expensive item in poultry diets because of the amount required (Olomu, 2011). The nitrogen corrected through metabolizable energy (TMEn) of corn is 3350 kcal/kg compared to 3300 to 3450 kcal/kg

for pearl millet (Prandini *et al.*, 2016). Thus, it is imperative to search for suitable alternative sources of energy in poultry feeds to reduce the cost of production. Corn can provide up to 65% metabolizable energy and 20% protein in poultry diets due to its high dietary inclusion rate (Chrysta *et al.*, 2020).

In Ghana, maize (both yellow and white) constitutes approximately 60 % of poultry feed formulations (Andam *et al.*, 2017). When feeding poultry, maize grains are either fed directly or ground and formulated with other suitable feed ingredients. The compounded feed thereafter fed to or transformed into the feed forms preferred by specific livestock species (Dei, 2017). The profitability and growth potential of the poultry industry in Ghana is significantly influenced by the availability and pricing of maize (Dela Cruz *et al.*, 2014). In order to reduce the cost of production and bolster the competitiveness of the local poultry industry, technological and innovative agronomic strategies need to be employed (Murdia *et al.*, 2016).

## **2.2 Importance of Maize in the Poultry Industry**

The preference for maize in poultry feed formulation stems from its accessibility (Shah *et al.*, 2016). Maize contains a vital raw material that has a higher calorific value, more amino acids, and fewer toxins than grains like broken rice and millet (Prandini *et al.*, 2016; Andam *et al.*, 2017). For many decades, maize has been a staple food for people. However, in certain jurisdictions around the world, its direct food consumption has decreased over the past few decades due to a variety of factors, including changes in eating patterns and rising income levels (Erenstein *et al.*, 2022). At the same time, maize has become more widely used in industrial applications and as feed for poultry. When utilized at 30 % or more in

the diet, yellow kernelled cultivars of maize are preferred as poultry feed because they are a strong source of  $\beta$ -carotenes and xanthophylls that give the skin, poultry fat, and egg yolk their yellow colour (Kaul *et al.*, 2019). Feed production in the poultry industry is the largest end-user of all cultivated maize (Afolayan *et al.*, 2015). Because of the increasing population and growing preference for higher protein intake from meat and eggs, there will likely be a greater need for and demand for maize farming. The relative cheap price and nutritional value of maize make it a more popular choice for poultry feed as compared to wheat and rice (Afolayan *et al.*, 2015).

### **2.3 Maize as a major feed ingredient**

The maize grain is the most significant in terms of poultry feed production or formulation. The other vegetative parts such as stalks, leaves, and young ears are used as forage or fodder for feeding ruminant livestock (Prandini *et al.*, 2016). It is known that among the cereal grains, maize has the highest conversion rate of dry matter into animal products like meat, eggs and milk (Jain *et al.*, 2016). The primary source of calories for feeding cattle, pigs, and poultry is maize (Aardsma *et al.*, 2017). The belief that maize has a steady and high nutritional content is one factor contributing to its widespread use in the diets of farmed cattle. Nevertheless, research conducted by Schedle (2016) has shown that maize's nutritional content and chemical composition vary to a greater extent, which makes the generic matrix values for the grain unreliable. The amount of starch, oil, protein, and anti-nutrients in maize determines its nutritional value for poultry (Erdaw *et al.*, 2016).

In Ghana and numerous other countries, maize is considered an essential cereal grain for monogastric farm animals, where it constitutes between 50 % and 60 % of their diets (Manu *et al.*, 2015). It is the main constituent in the majority of pig and poultry diets. Maize is highly palatable, easy to digest, low in fibre, and high in energy. The two main drawbacks of the typical maize cultivars grown in Ghana and other countries are their low protein content (9–10%) and their deficiency in certain essential amino acids, especially tryptophan and lysine (Badu-Apraku and Fakorede, 2017). As a result, it is not a sufficient supply of protein for monogastric. Normal maize contains about 10 % protein, but monogastric animals, including humans, cannot consume this because it lacks two necessary amino acids, tryptophan and lysine (Olabode *et al.*, 2021). To achieve a balanced nutrition in poultry production, it necessitates the use of highly rich protein supplements or synthetic amino acids such as lysine in poultry diets containing relatively large proportion of maize (Manu *et al.*, 2015).

## **2.4 Nutrient Composition of Maize Grain**

The nutritional composition of maize includes not only carbohydrates but also essential nutrients like crude protein and phosphorus (Rouf Shah *et al.*, 2016). It contains 7.5 %, 3.5 %, 1.9 %, 1.1 % and 3,373 kcal/kg of crude protein, crude fat, crude fibre, ash and metabolizable energy respectively (Rouf Shah *et al.*, 2016). Several factors affect the nutritional composition and value of maize. This includes but not limited to environmental conditions, post-harvest handling and genetic variability (Enyisi *et al.*, 2014). Environmental conditions such as edaphic quality, cultivation practices and climate influence nutrient content. Although maize can grow on varied soil types and conditions; however, the best soil type for producing maize with higher nutritional content is well

managed loamy because it is rich in essential plant nutrients particularly potassium (K), nitrogen (K), sulfur (S), phosphorus (P) and zinc (Zn) which are necessary for the production of energy dense and protein rich maize (Enyisi *et al.*, 2014). Storage conditions and methods of processing are post-harvest handling that can affect the preservation of nutrients in maize. Genetically, different cultivars of maize have varying nutrient compositions. The quality protein maize varieties (Obaatanpa) have proportionally higher nutritional values as compared to the conventional maize varieties (Accra Local) (Prandini *et al.*, 2016; Badu-Apraku and Fakorede, 2017). In terms of crude protein content, the quality protein maize varieties (Obaatanpa) contain 9.5-10.5 % while conventional maize varieties (Accra Local) contain 8.6 -9.5 % (Mwambo *et al.*, 2020).

## **2.5 Energy Content of Maize Grain**

Maize has been recognized worldwide as a major energy feed ingredient in the diets of poultry due to its high carbohydrate content and palatability (Jain *et al.*, 2016). Maize has a higher metabolizable energy value (3,365 Kcal/kg) (Afolayan *et al.*, 2015) than other potential poultry feed sources, such as rice (3,320 Kcal/kg), rice bran (2,620 Kcal/kg), peanuts (2,915 Kcal/kg), and oilcake (2,350 Kcal/kg) and as a result it is frequently used as a standard for measuring other energy feed sources (Andam *et al.*, 2017; Aardsma *et al.*, 2017). In poultry feed industry, maize which constitute a greater percent of the diet, is the most often used energy source (Dei, 2017). Corn is the most popular because of its easy availability and higher energy content. The feed and poultry industry standard for energy requirements in chickens is 3,200 kcal/kg for broilers and 2,600 kcal/kg for layer feed (Hellin *et al.*, 2013).

## 2.6 Protein Content of Maize Grain

The protein content of the maize grain is low, with a standard error of about 7 g/kg of crude protein (Barros *et al.*, 2017). Eight (8) to eleven (11) g of protein per 100 g of dry matter are found in maize grain. The amount of protein in the different grain fractions varies greatly. The germ has significantly more protein (184 g/kg DM) than the endosperm (80 g/kg DM), although the endosperm contains the majority of the grain's protein (Dei, 2017). In general, the grain's low protein content typically limits its nutritional value for both human and animals (Barros *et al.*, 2017). Each protein fraction's amino acid content and the relative proportions of the different protein fractions determine the overall maize grain's amino acid composition (Gebru *et al.*, 2019).

The proteins found in maize grain endosperm are commonly known as albumins, globulins, prolamins, and glutelins, depending on which solvent system they dissolve in. Glutamins and prolamins, commonly known as storage proteins, are only found in the endosperm, while albumins and globulins, generally known as water-soluble proteins, are also present in the germ and the aleurone layer (Ortiz-Martinez *et al.*, 2017). Approximately 50–60% of the total protein in typical maize grain is prolamin, which has a higher proportion than glutelin (Chen *et al.*, 2018). The relative proportion of each protein fraction has a significant impact on the amount of each individual amino acid in the total grain protein. Each protein fraction typically has a distinctive amino acid composition. Because lysine is the most deficient prolamin, maize protein has a low nutritional value (Ortiz-Martinez *et al.*, 2017; Chen *et al.*, 2018). In essence, the overall lysine deficiency in maize grain results from its low albumin and globulin content, which, in addition to having a high lysine content, shows a well-balanced amino acid composition comparable to that of animal

proteins with greater nutritional value (Ahmed *et al.*, 2023). Additionally, maize prolamins have higher levels of leucine than isoleucine, which results in the common amino acid imbalance that lowers maize's protein quality (Gebru *et al.*, 2019).

## **2.7 Non-Conventional Feed Materials for the Study**

The rising cost of conventional feed ingredients, such as maize and soya beans, has made poultry production more expensive, particularly for small and medium-scale farmers. As a result, there is growing interest in utilizing non-conventional feed ingredients that are locally available and often more affordable. Non-conventional feeds refer to alternative ingredients such as agro-industrial by-products, leaves, and other plant materials not traditionally used in poultry diets. These feeds have the potential to reduce feed costs, enhance sustainability, and support the growth of poultry in resource-limited settings (Tenza *et al.*, 2024). However, the use of non-conventional feeds also poses certain challenges regarding nutrient availability, palatability, and potential anti-nutritional factors (ANFs).

Studies have shown that non-conventional feed ingredients can have varying effects on the growth performance of poultry, depending on their nutrient composition and inclusion levels. For example, cassava peels, which are high in fibre but low in protein, may reduce growth rates if included at high levels without proper supplementation (Adeyemo *et al.*, 2020). However, when supplemented with protein-rich ingredients such as moringa or soya bean meal, cassava peels can be effectively utilized in poultry diets without negatively affecting performance (Ogbuewu *et al.*, 2023).

One of the main challenges associated with non-conventional feeds is the presence of ANFs, which can inhibit nutrient absorption and reduce the overall performance of poultry. For example, cassava peels contain cyanogenic glycosides, which can be toxic if not properly processed (Panghal *et al.*, 2021). Similarly, Leucaena leaves contain mimosine, an alkaloid that can cause toxicity in poultry if consumed in large quantities (Ramli *et al.*, 2017). Proper processing methods such as drying, boiling, or fermentation can help reduce the levels of ANFs in non-conventional feed ingredients, making them safer for poultry consumption.

### **2.7.1 Exploring non-conventional feed resources for sustainable livestock production in developing countries**

The availability of feed resources and their rational utilization for livestock represents possibly the most compelling task facing planners and animal scientists in the world. The situation is acute in numerous developing countries where chronic annual feed deficits and increasing animal populations are common, thus making the problem a continuing saga (Ramli *et al.*, 2017).

Thus, non-conventional such as full fat or soya bean oil feeds could partly fill the gap in the feed supply, decrease competition for food between humans and animals, reduce feed cost, and contribute to self- sufficiency in nutrients from locally available feed sources (Mashiloane *et al.*,2025). It is therefore imperative to examine for cheaper non-conventional feed resources that can improve intake and digestibility. Feedstuffs such as fish offal, cassava peel, palm kernel cake, sugarcane bagasse, unrefine soya bean oil, refine soya bean oil, rice bran, maize bran, cocoa bean waste, coconut meal, corn cob, moringa

leaf, leucaena leaf, local brewery and distillery by-products, sisal waste, and coffee pulp are commonly used in Ghana and could be invaluable feed resources for small and medium size holders of livestock (Ramli *et al.*, 2017).

## **2.8 Soya Bean Production in Ghana**

The soya bean (*Glycine max*) is an annual herbaceous erect plant that ranges in height from 30 to 183 cm (Mishra *et al.*, 2024). It has fine brown or grey hair on its stem, leaves, and pods. Soya beans, like peas, beans, lentils, and peanuts, are members of the Leguminosae or Fabaceae family. Soya beans have about 8 % of seed coat or hull, 90 % of cotyledon and 2 % of germ (Mishra *et al.*, 2024). Originally from East Asia, it has spread throughout the world, with the United States, Brazil, and Argentina being the leading producers (Bankole, 2022). It is mainly used for vegetable oil and oilseed meal production for use in feeding animal. The key factor boosting soya bean production has been the increase in the use of soya bean as a protein substitute for animal protein diets (Mawiya, 2016). The major farmed animal species diets containing soya bean include poultry, pigs, cattle and aquatic. In some animal feeds, soya bean is utilized comparatively more than in others. The primary goal is to supply pigs and poultry with high-quality protein (Mawiya, 2016). Out of all the plant protein sources, the majority of the land required for production of animal product is devoted to the cultivation of soya bean (Thrane *et al.*, 2017). For instance, the United States of America (USA) and the European Union (EU) both utilize a lot of soya bean in animal feed. Soya bean acreage of 5.0 million hectares in Brazil and 4.2 million hectares in Argentina is required for the EU's yearly livestock consumption alone. Therefore, because of its high protein content, energy contribution, and physiological advantages, soya beans continue to be essential in poultry nutrition (Thrane *et al.*, 2017). It provides the main

sources of high-quality protein and energy and is essential for maximizing feed efficiency, growth performance, and poultry health in general (Mawiya, 2016).

### **2.8.1 Nutrient composition of soya bean**

In broiler chickens, soya beans are regarded as a great source of protein and oil (Bankole, 2022). They include 20 % oil and 38 % crude protein as in Table 2.1. As a result, they are regarded as the most cost-effective since they eliminate the need for extra oils in the diet (Muslyumova *et al.*, 2021). As the most popular vegetable protein in animal feed, soya beans are a great feed substitute for animal proteins due to their high protein content (44–48 %) and balanced amino acid profile. However, raw soya bean meal contains anti-nutritional factors that reduce its digestibility and utilization (Alagawany *et al.*, 2019). Animal performance is adversely affected by these anti-nutritional compounds, which include non-starch oligosaccharides, polysaccharides, lectins, tannins, saponins, phytate, protease inhibitors, and antivitamin (Nabizadeh *et al.*, 2018). Soya beans need to be heated and under high pressure to lessen the effects of these chemicals, but heat treatment must be done carefully to prevent protein and other vital components from being denatured (Avedeweh, 2015).

**Table 2. 1: Nutrient Composition of Soya Bean Grain**

Parameter	(%)
Moisture content	9.000
Crude fat	15.50
Crude protein	43.20
Crude fibre	16.77
Carbohydrate	9.530
Ash content	6.000

Source: Avedeweh (2015)

### **2.8.2 Crude Protein and Amino Acids Content in Soya Bean**

According to Muslyumova *et al.* (2021), the soya bean seed has the highest crude protein content (about 40%) and the best amino acids among all the legume seeds. According to Thrane *et al.* (2017), soya bean protein contains all the essential amino acids required to meet nutritional needs of poultry for development, maintenance, and growth. Protein serves as the building block for organs, muscle, feathers, and eggs (Alagawany *et al.*, 2019). He *et al.* (2021) claimed that birds' daily diets must contain ten amino acids; however, this contradicts report of Classen (2017) who opined that pigs and poultry nutrition require only five important amino acids, namely tryptophan, cysteine, theronine, methionine, and lysine. Soya bean has the highest lysine digestibility (91%) (Classen, 2017); it also high in methionine, cysteine, and theronine digestibility (He *et al.*, 2021).

## **Carbohydrate content of soya bean**

In addition to being high in protein, soya beans are also high in carbohydrates (Al Loman *et al.*, 2016). Poultry use glucose, which is produced from carbohydrates, as an energy source for development and egg production. Adebowale *et al.* (2019) clarified that monogastric animals have poor digestion of the carbohydrate components (hulls, sugars, and non-starch polysaccharides). About 10% of the carbohydrates in soya beans are free sugars (5% sucrose, 4% stchyyose, and 1% Raffi nose) (Al Loman *et al.*, 2016; Nguyen *et al.*, 2021). Since monogastric animals lack the enzymes necessary to hydrolyze these carbohydrates, bacterial fermentation is used to digest these fibres (Adebowale *et al.*, 2019). Soluble non-starch polysaccharides (polymers that are partially soluble in water) and insoluble non-starch 13 polysaccharides (cellulose) are the two primary categories of non-starch polysaccharides (Nguyen *et al.*, 2021). In chickens, insoluble non-starch polysaccharides (cellulose) can impair growth performance and decrease nutrient digestibility. Soya bean meals must have 35–40% carbohydrates when oil is produced (Choct, 2018).

### **2.8.3 Crude Fat Content of Soya Bean**

One of the main sources of nutritional energy in poultry feed is crude fat, also known as ether extract. The amount of crude fat in soya beans might vary greatly according to the processing method used. Full-fat soya beans (FFSB) are a high-energy feed item because they normally contain 18–20 % crude fat (Tang *et al.*, 2024). However, as most of the oil is eliminated during processing, solvent-extracted soya bean meal (SE SBM), the most popular type in commercial chicken feeds, has as little as 0.5–1 % crude fat (Kerr *et al.*, 2023). A compromise between cost and energy contribution is provided by expeller-

processed soya bean meal, which preserves roughly 6–7 % fat through mechanical extraction as opposed to chemical extraction (Kerr *et al.*, 2023).

In broilers and layers, it has been demonstrated that adding high-fat soya bean products, like FF SB, improves energy density and feed conversion. However, it is important to balance the amount of protein and energy in the diet. Overconsumption of fat can result in increased deposition of fat in the abdomen, which is undesirable for carcass quality, particularly in broiler chickens (Ali *et al.*, 2022). The significance of regulated inclusion levels was highlighted by a study by Al-Marzooqi and Leeson (2016), which showed that diets heavy in soya bean oil or fat enhanced energy utilization but also increased fat pad weights.

The unsaturated fatty acid fat found in soya beans mostly improves meat quality by increasing tenderness and lowering the amount of saturated fat in poultry meat (Wang *et al.*, 2017). As a result, the fat content of soya bean products does not influence growth performance of poultry but also affect the nutritional value of poultry products intended for human consumption.

#### **2.8.4 Crude Fibre Content of Soya Bean**

Crude fibre is a measurement of the amount of indigestible cellulose, pentosans, lignin, and other comparable substances present non feed (Slominski, 2018). Crude fibre is essential for gut health and digestive efficiency, despite being frequently viewed as a less desirable part of chicken diets because of the birds' poor capacity to digest fibrous materials. Processing also affects the amount of crude fiber in soya bean products. While non-dehulled or hull-containing soya bean meals may include up to 7–14 % crude fibre,

dehulled soya bean meal can have as little as 3–5 % (Okonkwo *et al.*, 2023). Unless added in very small amounts or processed with enzymes, soya bean hulls a by-product of processing can contain more than 50 % crude fibre (Adewole *et al.*, 2016), making them an unsuitable option for monogastric animals like chicken.

For instance, diets with slightly higher levels of dietary fiber from soya bean hulls or full-fat soya bean by-products were linked to increased intestinal villus height and improved nutrient absorption in broilers (Singh *et al.*, 2018). However, excessive fiber, especially when undigested, can reduce nutrient availability and feed efficiency, resulting in lower growth rates and increased feed costs. Even though poultry lack the complex digestive systems necessary to ferment fiber efficiently, moderate inclusion of dietary fiber has been shown to promote intestinal development, gizzard function, and beneficial microbiota populations (Jiménez-Moreno *et al.*, 2019). This means that when adding high-fibre soya bean products to chicken diets, fibre-degrading enzymes like xylanase and cellulase must be used. It has been demonstrated that this type of enzyme supplementation enhances the use of fibrous components, especially in feeds formulated from soya bean (Masey O'Neill *et al.*, 2020).

### **2.8.5 Mineral Content of Soya Bean**

Poultry's nutritional needs for minerals are largely met by dietary soya bean. Calcium (Ca), phosphorus (P), potassium (K), magnesium (Mg), and sulfur (S) are among the essential macro and microminerals found in it. Soya beans contain roughly 0.20–0.25 % calcium and 0.59–0.65 % total phosphorus, although only 0.20–0.21 % of this is accessible to chicken because of the high phytate-bound proportion, according to Olukosi *et al.* (2017).

Magnesium and sulfur are present at 0.21–0.27 % and 0.30–0.43 %, respectively, while potassium level ranges from 1.7 to 1.97 %. As stated in table 2.2. There are also appreciable concentrations of microminerals including zinc (35–60 ppm), copper (15–28 ppm), iron (75–160 ppm), and manganese (27.5–32.3 ppm) (Lee *et al.*, 2022).

Because phytate chelates and phosphorus make it unavailable to monogastric animals, phosphorus digestibility is a major concern in poultry nutrition (Bougouin *et al.*, 2015). The necessity for standardization was highlighted by Rojo *et al.* (2023), who discovered that the phosphorus digestibility in SBM varied from 35.3 % to 63.2 % based on the soya bean source and processing. Phosphorus availability has been demonstrated to be enhanced by autoclaving and enzymatic treatments, especially the addition of microbial phytase (Walk *et al.*, 2021). Studies have shown that supplementing soya bean with phytase improved the digestibility of ileal phosphorus from 46 % to 61 %, with higher enzyme dosages achieving up to 77 % digestibility (Adeola and Cowieson, 2016; Walk *et al.*, 2021). In a similar vein, autoclaving soya bean has been shown to increase phosphorus bioavailability by breaking up phytate complexes (Ravindran *et al.*, 2017).

**Table 2. 2: Mineral Composition of Soya Beans**

<b>Mineral</b>	<b>Composition</b>
Calcium (Ca) (%)	0.200–0.250
Potassium (K) (%)	1.700–1.970
Sodium (Na) (%)	0.040
Phosphorus (P) (%)	0.590–0.650
Magnesium (Mg) (%)	0.210–0.270
Chloride (Cl) (%)	0.020–0.030
Sulfur (S) (%)	0.300–0.430
Chloride (Cl) (%)	0.020–0.030
Copper (Cu) (ppm)	1500–28.00
Iron (Fe) (ppm)	75.00–160.0
Selenium (Se) (ppm)	0.100
Zinc (Zn) (ppm)	35.00–60.00
Manganese (Mn) (ppm)	27.50–32.30

**Source: (Lee *et al.*, 2022; Olukosi *et al.*, 2016)**

### **2.8.6 Vitamin Composition of Soya Bean**

Soya bean contains a variety of vitamins that promote the poultry's physiological processes as in table 2.3. These include water-soluble B-complex vitamins and fat-soluble vitamins like vitamin E. Vitamin E, an essential antioxidant that promotes immune system function and preserves cellular integrity, is found in soy beans (Surai, 2016). According to Nguyen *et al.* (2021), soya beans typically contain 3.0–6.6 mg/kg of vitamin E. The most prevalent B-vitamins are pantothenic acid (13.2–13.8 mg/kg), thiamin (1.7 mg/kg), riboflavin (2.6–

4.4 mg/kg), and niacin (20.9–59.8 mg/kg). Choline, which is essential for lipid metabolism and liver function in poultry, is present at roughly 2673–2850 mg/kg (Olukosi and Adeola, 2017), while biotin and folic acid are detected in the range of 32.0–320 µg/kg and 450–700 µg/kg, respectively (Batal and Dale, 2016).

The processing techniques, however, can have a significant impact on the vitamin concentration and bioavailability. Trypsin inhibitors and other anti-nutritional factors must be deactivated by heat treatment, which can also degrade heat-sensitive vitamins like folic acid and thiamin (Krautwald-Junghanns *et al.*, 2022). Overheating decreased the levels of riboflavin and vitamin E, which may compromise the nutritional value of soya bean (Nguyen *et al.*, 2021). Therefore, maintaining the soya bean's vitamin integrity requires optimizing processing techniques.

**Table 2. 3: Vitamin Composition of Soya Bean**

<b>Vitamin</b>	<b>Composition</b>
Vitamin E (mg/kg)	3.000–6.600
Thiamin (B1) (mg/kg)	1.700
Riboflavin (B2) (mg/kg)	2.600–4.400
Niacin (Vitamin B3) (mg/kg)	20.90–59.80
Pantothenic Acid (B5) (mg/kg)	13.20–13.80
Biotin (B7) (µg/kg)	32.00–320.0
Folic Acid (Vitamin B9) (µg/kg)	450.0–700.0
Choline (mg/kg)	2673–2850

**Source: (Surai, 2016; Nguyen *et al.*, 2021)**

### 2.8.7 Full-fat soya bean (FFSB) Processing and its Importance

Full fat soya bean are complete soya beans in which the oil has not been extracted. The product is made using a number of techniques, including dry or wet extrusion, frying, autoclaving, roasting, and micronizing to deactivate the ANFs. Raw soya bean contain anti nutritional factors like trypsin inhibitors, phytates and lectins which can impair nutrient utilization and cause digestive issues (Das, 2019). Various methods can reduce ANTs including heat method and enzyme treatment. Processing soya bean improve protein digestion, increase energy availability and reduce ANTs. Depending on the extent of heat damage or ANF inactivation, each of these procedures has a unique effect on the soya bean's nutritional value (NRC, 1994).

Full fat soya bean is processed by removing impurities and dry the beans to moisture content of 10-12% heat the raw soya beans to 248-266F for 20-30 minutes, stirring occasionally, to inactivate trypsin inhibitors. Allow the soya beans to cool then grind them into meal (Zhan et al., 2022).

**Table 2. 4: Nutritional Composition of Full Fat Soya Bean**

Parameter	Full fat soya bean
Dry Matter (%)	89.40
Crude Protein (%)	37.10
Crude Fibre (%)	5.100
Ether Ext ract (%)	18.40
Ash (%)	4.900
Gross Energy (MJ/kg)	20.95

**Source: Voss et al. (2018).**

### **2.8.8 Impact of Full-Fat Soya Bean Diet on the Growth Performance of Broilers**

Numerous studies confirm that including processed FFBS in broiler diets significantly improves body weight gain. Karunaratne *et al.* (2023) demonstrated that broilers fed diets containing up to 20 % extruded FFBS exhibited superior weight gains relative to birds on soya bean meal (SBM) and supplemental oil-based diets. The improvement was attributed to enhanced energy provision and superior protein quality. Alagawany *et al.* (2022) similarly reported increased average daily gains (ADG) in broilers fed 15 % extruded FFBS diets during both starter and grower phases. These enhancements in BWG are linked to better utilization of dietary nutrients when ANFs are adequately neutralized. Conversely, Raji *et al.* (2021) highlighted that raw or insufficiently processed FFBS negatively impacts BWG, mainly due to residual trypsin inhibitor activity interfering with protein digestion. Feed intake in broilers is influenced by diet palatability, nutrient density, and textural quality. Bello and Olomu (2023) found that the inclusion of extruded FFBS slightly increased feed intake in broilers due to improved pellet quality and reduced dustiness from inherent oil content. Similar findings were reported by El-Safty *et al.* (2022), who observed enhanced feed consumption in broilers fed diets containing 10-15 % FFBS. However, excessive inclusion of poorly processed FFBS may depress feed intake due to bitterness from residual anti-nutritional compounds (Mabelebele *et al.*, 2021). Thus, optimal inclusion levels combined with adequate processing are essential to maximize palatability and intake.

Feed conversion ratio is a pivotal parameter reflecting the efficiency of feed utilization for body weight gain. Several researchers have noted that broilers fed FFSB-based diets demonstrate improved FCR compared to conventional SBM diets supplemented with oil (Bello and Olomu, 2023). Karunaratne *et al.* (2023) observed that inclusion of extruded FFSB at 15-20 % in broiler diets consistently yielded better FCR values, indicating more efficient nutrient utilization. This efficiency is attributed to the readily available dietary lipids from FFSB, which reduce metabolic energy expenditures associated with digestion (Alagawany *et al.*, 2022). Nevertheless, inclusion rates exceeding optimal levels or inadequate thermal processing can compromise FCR due to impaired nutrient availability and increased energy demand for detoxification of ANFs (Raji *et al.*, 2021).

Processing full-fat soya beans is critical in enhancing their nutritional value for broiler diets. Among various methods, extrusion is the most effective, as it efficiently inactivates ANFs while preserving nutrient integrity (Ravindran, 2023). Studies by Mabelebele *et al.* (2021) indicated that broilers fed extruded FFSB exhibited higher apparent metabolizable energy (AME) and protein digestibility, culminating in improved BWG and FCR. Autoclaving and roasting are also effective, but extrusion remains superior in preserving oil integrity while neutralizing ANFs (Bello and Olomu, 2023). Raji *et al.* (2021) demonstrated that improper heat treatment led to significant reductions in BWG and poorer FCR in broilers.

The impact of FFSB inclusion varies across different growth phases in broilers. Adeyemo *et al.* (2020) emphasized that starter-phase broilers benefit most from FFSB inclusion, particularly due to higher energy requirements during early rapid growth stages. Inclusion

rates of 10-15% were found to be optimal for achieving significant BWG improvements in young chicks (El-Safty *et al.*, 2022). During grower and finisher phases, birds exhibit tolerance to higher FFBSB inclusion levels (up to 20 %), provided the diets are isonitrogenous and supplemented with limiting amino acids (Karunaratne *et al.*, 2023). Gradual incorporation of FFBSB, with concurrent amino acid fortification, maximizes its benefits throughout the production cycle.

Comparative evaluations highlight the efficacy of FFBSB relative to conventional SBM and supplemental oil diets. Bello and Olomu (2023) noted that broilers fed FFBSB-based diets not only exhibited comparable or superior growth metrics but also benefited from improved feed homogeneity and potentially lower formulation costs. Similarly, Alagawany *et al.* (2022) emphasized that substituting SBM plus oil with FFBSB simplifies diet formulation and reduces ingredient variability. However, attention must be paid to amino acid balance, especially regarding methionine and lysine levels (Karunaratne *et al.*, 2023).

### **2.8.9 Soya Bean Oil Production and Uses**

Soya bean oil is produced primarily for human consumption. However, it has become a useful source of feed-grade fat for animals due to a need to formulate high-energy diets for modern breeds. Feed-grade soya bean oil is popularly used in high energy diets, particularly for poultry, because of its high digestibility and metabolizable energy content compared with other vegetable fats/oils (Bergeron, 2021). It is used widely in rations for broiler chickens and growing turkeys as a feed-grade fat to increase energy density of feeds and improve efficiency of feed utilisation (Babatunde *et al.*, 2021). The high energy value of soya bean oil is attributed to its high percentage of (poly) unsaturated fatty acids, which

are well absorbed and utilised as a source of energy by the animal (Castro *et al.*, 2019). Also, the high polyunsaturated fatty acids (PUFA) in soya bean oil appears to have an energy independent effect on improving reproduction in dairy cattle (Rahbar *et al.*, 2014), and this has been attributed to the role of linoleic acid in reproduction (Szczyko *et al.*, 2020).

## **2.9 Use of Fats and Oils in Poultry Diets**

Fats and oils are normally used as an energy source in poultry diets due to its high energy density. Fat energy is at least twice that of protein and carbohydrates (NRC, 1994), and metabolizable energy values are around 37.7 kJ for 1 gram of fat, 16.7 kJ for 1 gram of protein and 16.7 kJ for 1 gram of carbohydrates (Singh *et al.*, 2017). Lipid addition has other benefits including a reduction in dustiness and binding of other nutrients, improved palatability and lubrication of equipment during preparation of food which improves pellet quality (Baião and Lara, 2005). Fat-supplemented diets increase feed efficiency and profitability in poultry (Sahito *et al.*, 2012). Besides, oil improves the palatability of diets, reduces the dustiness of feeds and decreases the passage rate of feed through the intestinal tract of poultry, which gives more time for the adequate absorption of all nutrients present in the diet (Baião and Lara, 2005).

The inclusion of fat and oil is a common practice in modern poultry production to increase the Fat and oil are commonly used in poultry diets to increase the energy density as they yield 2.25 times more calories than carbohydrates and protein energy content of diet. The addition of fat to diets, besides supplying energy, improves the absorption of fat-soluble vitamins, diminishes the pulverulence, increases the palatability of the rations, and

increases the efficiency of the consumed energy (lower caloric increment) (Olabode *et al* 2021). Furthermore, it reduces the passage rate of the digesta in the gastrointestinal tract, which allows a better absorption of all nutrients present in the diet. High energy diets have been shown to improve growth and feed efficiency (Lammers *et al.*,2000). Oils added to the rations of animals are effective on the fatty acid composition and amount of abdominal fat. In fact, fatty acids composition of oils used in poultry rations are reflected in the animal products because dietary fatty acids are incorporated with little change into the bird body fats (Williams *et al.*, 2022. Thus, the type of fat used in the feed influence the composition of broiler body lipids. Abdominal fat is a good indicator of chicken body fats because it is very sensitive to changes in dietary fatty acid composition. Crespo *et al.* (2001) have reported that broiler chickens fed with diets enriched of polyunsaturated fatty acids have less abdominal fat or total body fat deposition than do broiler chickens fed with diets containing saturated fatty acids.

Barbour (2006) reported that by increasing the proportion of supplemental animal-vegetable blend fat from 0 to 60 g/kg in isocaloric diets (2,963 kcal/kg), feed conversion was improved without any effects on BW, feed intake, abdominal fat, or whole carcass composition. In a subsequent experiment, 30, 60, or 90 g/kg of corn oil added to isocaloric corn-soya bean meal diets (2,926 kcal/kg) increased BW in four weeks old male broilers compared with those fed equal quantities of diets with no added oil. Those improvements in performance were related to enhanced use of calories beyond what was accounted for in terms of calculated dietary ME, and this was especially true in the case of vegetable oils. In more recent studies, linear increases in weight gain and feed efficiency were observed when male broiler chickens were fed corn-wheat-soya bean meal diets in which ME was

increased from 2,800 to 3,000 kcal/kg through changes in supplemental soya bean oil or dietary carbohydrate levels (Williams *et al* 20220).

## **2.10 Effect of Soya Bean Meal on the Growth Performance of Broilers**

Soya bean meal (SBM) is a widely used protein source in poultry diets due to its high protein content, balanced amino acid profile, and digestibility (Abdel-Raheem *et al.*, 2023). It is obtained as a by-product of soya bean oil extraction and serves as an economical alternative to fishmeal and other protein sources in broiler diets. The nutritional value of SBM is influenced by processing methods, which determine its amino acid availability and overall quality (Asghar *et al.*, 2024). Its inclusion in broiler diets has been extensively studied for its impact on growth performance, including body weight gain, feed intake, and feed conversion efficiency.

Several studies have demonstrated that broilers fed diets containing SBM exhibit improved growth performance compared to those on diets with lower-quality protein sources. Olukosi *et al.* (2017) found that replacing fishmeal with SBM resulted in comparable weight gain and feed conversion ratios (FCR) in broilers. This is attributed to SBM's high crude protein content (44-48%) and favourable amino acid composition, particularly lysine and methionine, which are essential for muscle growth and development. The digestibility of SBM is another critical factor influencing broiler growth. According to Anyiam *et al.* (2025), the digestibility of amino acids in SBM is superior to other plant-based protein sources, leading to better nutrient utilization. However, the presence of anti-nutritional factors such as trypsin inhibitors and lectins may reduce protein digestibility if not properly processed (Thakur *et al.*, 2019). Heat treatment and enzyme supplementation have been

shown to enhance the digestibility and efficiency of SBM-based diets, ultimately improving broiler performance.

SBM's impact on feed intake is also well-documented. Broilers tend to consume diets containing SBM more readily than those with lower digestibility protein sources, such as sunflower or cottonseed meal (Amerah,*et al*, 2015). This is likely due to its palatability and amino acid balance, which meet the nutritional requirements of growing broilers. Higher feed intake, coupled with improved nutrient utilization, results in better overall growth performance. The effect of SBOM on the feed conversion ratio (FCR) is significant in poultry nutrition research. Studies by Wu *et al.* (2019) indicate that broilers fed SBM-based diets achieve lower FCR values, meaning they convert feed into body mass more efficiently than those on alternative protein sources. This improved efficiency can be attributed to SBM's highly digestible protein and essential amino acids, which optimize metabolic functions and muscle accretion.

Economic considerations also play a crucial role in the inclusion of SBM in broiler diets. Given its relatively low cost compared to animal protein sources, SBM provides a cost-effective means of meeting broilers' protein needs while maintaining optimal growth rates (Amer *et al.*, 2021). Moreover, the global availability of SBM makes it a sustainable choice for poultry producers seeking to reduce feed costs without compromising bird performance. The effect of SBM on carcass traits is another aspect of interest. Research by Hussein *et al.* (2020) suggested that broilers fed SBM-based diets exhibit improved carcass yield, particularly in breast muscle development. Additionally, the fat content of broiler

meat may be influenced by dietary SBM levels, with some studies reporting lower abdominal fat deposition in birds fed higher levels of SBM (Katoch *et al.*, 2022).

Despite its numerous benefits, SBM inclusion in broiler diets must be optimized to prevent nutrient imbalances. Excessive dietary SBM levels can lead to amino acid imbalances and suboptimal growth performance (Ravindran *et al.*, 2017). Therefore, proper formulation and supplementation with synthetic amino acids, such as methionine and lysine, are recommended to ensure an ideal nutrient profile for broilers. Recent advancements in feed technology have explored the combination of SBM with feed additives like probiotics, enzymes, and organic acids to enhance its nutritional value further (Zhang *et al.*, 2022). These additives help improve nutrient absorption, reduce the impact of anti-nutritional factors, and optimize growth performance.

## **2.11 Effect of Soya Bean Oil on Carcass Traits in Broilers**

Soya bean oil is widely used as an energy-rich lipid source in broiler diets. Its inclusion in poultry feed has shown significant effects on carcass traits, including dressing percentage, fat deposition, and meat yield. These effects are mainly attributed to its high energy density and essential fatty acid content, especially linoleic acid.

According to Afolayan *et al.* (2015), dietary inclusion of soya bean oil at 4–6% significantly improved carcass yield and breast muscle weight in broilers. However, higher levels led to increased abdominal fat deposition, indicating the importance of balancing energy levels in feed formulation. Similarly, Arteaga-Wences *et al.* (2021) reported that soya bean oil enhanced feed efficiency and promoted better muscle-to-fat ratios in broiler carcasses.

One of the primary effects of SBO on broiler carcass traits is its influence on carcass yield. Studies have shown that broilers fed diets with optimal levels of SBO exhibit higher dressing percentages compared to those on alternative protein sources such as sunflower or cottonseed meal (Ven der Merve *et al.*, 2019). This can be attributed to SBO's high lysine content, which plays a crucial role in muscle growth and protein deposition (Olukosi *et al.*, 2017). Properly balanced SBO inclusion ensures that birds reach market weight with minimal carcass waste.

Breast muscle development is another critical aspect of carcass traits affected by SBO. As a rich source of fatty acid, SBO enhances calories synthesis, leading to improved breast meat yield and fat pad Research by Wu *et al.* (2018) found that broilers receiving SBO-based diets had significantly larger breast muscles compared to those fed lower-quality protein sources. Since breast meat is a highly valuable portion of the broiler carcass, optimizing SBO inclusion in diets enhances profitability in poultry production. The impact of SBO on abdominal fat deposition is also well-documented. While protein sources influence muscle growth, they can also affect fat accumulation. Studies indicate that broilers fed high levels of SBO tend to have high abdominal fat compared to those receiving diets with lower-quality protein sources (Gilbert, 2011). This is attributed to SBOs favourable energy-protein ratio, which increases excessive fat deposition while promoting lean muscle growth (Choi *et al.*, 2023). Lower abdominal fat levels are desirable in broiler production, as they improve meat quality and consumer preference.

SBO also influences the composition of intramuscular fat, which affects meat texture and juiciness. A study by Ravindran *et al.* (2017) found that broilers on SBO-based diets had

moderate intramuscular fat levels, enhancing meat tenderness without excessive fat accumulation. The balance between protein and energy in SBO diets plays a crucial role in optimizing these traits, ensuring that broiler meat remains both nutritious and palatable. Proper protein nutrition, including fatty acid from SBO, supports skeletal growth and bone mineralization (De Coca-Sinova *et al.*, 2017). Strong skeletal structure is essential for broilers to support rapid weight gain and avoid processing defects such as broken bones during slaughter. Enzyme supplementation in SBO diets has been shown to improve calcium and phosphorus utilization, further enhancing bone integrity (Batal and Parsons, 2016).

Meat quality attributes such as colour, texture, and water-holding capacity are also influenced by SBO inclusion. Research by Zhang *et al.* (2022) indicates that SBO contributes to improved meat texture by enhancing protein deposition and reducing excessive connective tissue formation. Additionally, its high digestibility and amino acid balance support optimal muscle hydration, reducing drip loss and improving overall meat quality.

Excessive SBO levels without proper amino acid supplementation may lead to disproportionate muscle growth or increased carcass fat content (Ravindran *et al.*, 2017). Therefore, proper diet formulation, including the addition of synthetic amino acids such as lysine and methionine, ensures balanced nutrient intake for ideal carcass development. The economic implications of SBO on carcass traits are also noteworthy. Since broiler meat yield and quality directly influence market value, incorporating SBO effectively in poultry diets can enhance profitability (Tahmisbi *et al.*, 2024). The widespread availability and

cost-effectiveness of SBO make it a preferred choice for poultry producers seeking to maximize carcass yield while maintaining production efficiency.

## **2.12 Impact of Soya Bean Oil on Bone Traits of Broilers**

Soya bean oil (SBO), a rich source of unsaturated fatty acids, is frequently used in poultry diets to enhance energy density and improve nutrient absorption. Its inclusion in broiler chicken diets has shown notable effects on growth performance and bone development. Soya bean oil improves the absorption of fat-soluble vitamins, particularly vitamin D3, which plays a key role in calcium and phosphorus metabolism. These minerals are vital for proper bone formation and mineralization (Rafeeq *et al.*, 2020). Improved vitamin D3 bioavailability enhances tibia ash content and bone strength in broilers. The inclusion of soya bean oil in broiler diets has been associated with increased tibia breaking strength and bone density, especially when diets are balanced for calcium and phosphorus. According to Zhao *et al.* (2020), broilers fed diets containing up to 6% SBO showed improved tibia strength compared to those on fat-free diet.

SBO contributes to energy balance, which is critical for optimal growth and skeletal development. Adequate energy allows for efficient protein and mineral utilization, thus supporting stronger and well-structured bones (Qamar *et al.*, 2025). While soya bean oil itself does not harm bone traits, excessive inclusion without proper adjustment of other nutrients (especially minerals) can result in nutrient imbalances, potentially weakening bone quality. High energy intake without adequate calcium/phosphorus supplementation can reduce bone mineralization and increase the risk of skeletal disorders (Vorland, 2017).

One of the key factors influencing bone health in broilers is dietary protein quality and digestibility. SBO contains moderate levels of essential amino acids, particularly lysine and methionine, which play roles in collagen synthesis and bone matrix formation (Rath *et al.*, 2017). Studies suggest that broilers fed SBO-based diets may exhibit similar or slightly improved bone development compared to those on diets with lower-quality protein sources, depending on inclusion levels and amino acid balance (Liu *et al.*, 2019).

Calcium (Ca) and phosphorus (P) metabolism are central to bone mineralization, and SBO contains phytates that can bind phosphorus and reduce its bioavailability (Couce *et al.*, 2021). However, phytase enzyme supplementation has been shown to significantly enhance phosphorus digestibility in SBO-based diets, which can contribute to improved bone mineralization and skeletal strength (Cowieson *et al.*, 2017). Bone density is an important measure of skeletal health. Some evidence indicates that inclusion of properly balanced SBO in broiler diets may support tibia and femur mineralization, particularly when phosphorus and calcium are adequately supplied and digestibility is optimized (Salim *et al.*, 2022). Additionally, the mechanical properties of bones, such as breaking strength, are influenced by nutrient composition and digestibility. Research by Zhang *et al.* (2022) noted that broilers receiving diets with balanced amino acid profiles, including those based on SBO, had stronger tibias with enhanced resistance to fracture. Histological analyses also show that well-formulated SBO diets may support favorable bone microarchitecture, such as increased trabecular thickness and organized osteocyte structure (Kim *et al.*, 2021).

While SBOM offers numerous benefits to bone health, its impact may vary based on dietary formulation and nutrient interactions. Excessive inclusion of SBO in broiler diets can lead

to amino acid imbalances or reduced calcium absorption, potentially compromising skeletal integrity (Salim *et al.*, 2022). Therefore, appropriate dietary formulation, including the supplementation of essential minerals and enzymes, is necessary to maximize the benefits of SBO for bone traits.

Comparative studies between SBO and alternative protein sources, such as fish meal and meat meal, have shown that SBO can be equally effective in supporting bone health when properly balanced with minerals (Jian *et al.*, 2016). However, further research is needed to optimize SBO inclusion levels and evaluate its long-term effects on skeletal development.

### **2.13 Lipid Metabolism**

Lipids are a highly concentrated source of energy, with more than twice the energy density of carbohydrates (Chandel, 2021). The synthesis of fatty acids from non-carbohydrate sources is very limited in broilers, especially in adipose tissue. A significant portion (80–85%) of the fatty acids stored in adipose tissue are derived from plasma lipids, which originate from either the diet or the liver. The liver plays a critical role by processing and assembling lipids into lipoproteins, which are then secreted into the bloodstream (Lal, 2023). Broilers have a limited ability to efficiently utilize dietary fats, particularly at a young age. The product of carbohydrate metabolism, acetyl-CoA, can be converted into fatty acids and stored as lipids, demonstrating the close link between the two pathways (Moffett, 2020).

Changes in the levels of protein, carbohydrates, and lipids in the diet significantly impact metabolic processes and overall performance. Adding emulsifiers to feed can improve fat utilization and digestion in broilers, potentially lowering costs (Chandel, 2021). Young

birds have physiological limitations in their capacity to digest and use dietary fats effectively. Supplementing diets with nutrients like zinc can modulate lipid metabolism and improve performance, even under challenges like necrotic enteritis (Lal, 2023).

### **2.15 Maize Grains**

Maize (*Zea mays* L.) is a major staple food grain throughout the world, particularly in Africa, Latin America and Asia, and a major feedstuff in developed countries. The maize grain has many food (grain, flour, syrup, oil etc) and non-food usages (cosmetics, adhesives, paints, varnishes). Maize starch and oil are also major products (Ecocrop, 2010). The maize grain is a major feed grain and a standard component of livestock diets where it is used as a source of energy. Other grains are typically compared to maize when their nutritional value is estimated. Many by-products of maize processing for flour (hominy feed, bran, germs, oil meal), starch (corn gluten feed, corn gluten meal) and alcohol/biofuel industries (distillers' dried grains and soluble) can be fed to animals.

### **2.16 Nutrient Composition of Maize**

Maize grain is a globally significant cereal grain and a foundational food source, particularly in many parts of Africa, Latin America and Asia (Erenstein, 2022). The composition of various nutrients in maize are tabulated in Table 2.5.

**Table 2. 5: Nutrient Composition of Maize.**

<b>Nutrient</b>	<b>Quantity</b>
Weight	100g
Energy	1651.08 KJ
Calories	390.76 kcal
Fat	4.38g
Protein	9.807g
Carbohydrate	73.85g
Fibre	8.35g
Water	10.56g

**Source: Avedeweh (2015)**

## **2.14 Carbohydrate Metabolism**

Carbohydrate and lipid metabolism in broilers are central to energy production and growth, with carbohydrates (especially glucose) serving as the primary energy source, while lipids provide concentrated energy (Singh *et al.*, 2017). These metabolic pathways are interconnected, as products of glucose metabolism, such as acetyl-CoA, can be converted into lipids for storage. Broiler metabolism is optimized by ensuring adequate dietary carbohydrates for immediate energy and by managing lipid intake and utilization, as young birds have limitations in efficiently digesting and utilizing fats. Glucose is the main energy source, converted into pyruvate, which then enters the citric acid cycle to produce large amounts of ATP through oxidative phosphorylation (Chaudhry, 2018). Dietary carbohydrates are essential for maintaining blood glucose levels, which are crucial for

tissues that rely heavily on glucose. Low-carbohydrate diets can lead to low glucose levels, impacting performance and growth.

## **CHAPTER THREE: MATERIALS AND METHODS**

### **3.1 Location and Duration of the Study**

The study was conducted at the Poultry section of the Department of Animal Science Education of the Akenten Appiah - Menka University of Skills Training and Entrepreneurial Development, Asante Mampong campus. Asante Mampong lies in the Transitional zone between the Guinea Savannah Zone of the North and the Tropical Rain Forest of the South of Ghana along the Kumasi-Ejura Road. The study lasted for two months. The study started in January and ended in March, 2025.

### **3.2 Dietary Treatments and Experimental Design**

The composition of homogenized primary components (maize, wheat bran, soya bean meal, and fishmeal) was assessed and applied in creating four experimental diets (Table 3.1). The experimental design used for the experiment was completely randomized design (CRD). The treatment groups included: Treatment one (T1) – Conventional soya bean meal diet (Control diet) Treatment two (T2) – Full-fat Soya beans meal diet, Treatment three (T3) – Soya beans Oil diet, and Treatment four (T4) - Full-fat Soya bean meal + Conventional soya bean meal in starter and grower/finisher diets. The diets were formulated using the Concept 5-feed formulation program from Creative Formulation Concepts, LLC, based in Annapolis, MD. After formulation, the nutrient composition of the diets was determined through proximate chemical analysis using procedure according to AOAC (1990). Throughout the starter phase (d 0 to 28), grower- finisher phase (d28 to 56), water and diets were freely available *ad libitum*. In all phases, the diets were provided in mash form.

**Table 3. 1: Composition and Calculated Analysis of Experimental Diet**

<b>Ingredients</b>	<b>Starter Diet</b>				<b>Grower/Finisher Diet</b>			
	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>T4</b>	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>T4</b>
Maize grain	65	55	56.5	55.04	74	65	64.5	66.69
Soya bean meal	15	0	14	4	7.22	0	7.9	6
Wheat bran	1.49	18.31	8	8	2.52	5.5	8	7
Fish meal	16.28	17.16	17	14	14.42	9.5	14.5	12.57
Dicphosphate	0.98	0.8	0.8	0.8	0.65	0.9	0.7	1.1
Salt	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Oyster shell	0.24	0.39	0.4	0.53	0.2	0.3	0.23	0.35
Full-fat soya bean		17.34	0	17	0	17	0	12.57
Soya bean oil	0	0	2.17	0	0	0	2.5	0
Mineral Premix	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Total	100	100	100	100	100	100	100	100
<b>Calculated Nutrient</b>								
Protein	23	23	23.09	23.08	20.0	20.0	20.0	20.0
Calcium	1	1	1	1	0.87	0.84	0.84	0.82
Fat	4.21	7.05	6.35	6.74	4.65	8.5	6.76	7.10
Phosphorus	0.45	0.48	0.47	0.45	0.34	0.35	0.38	0.4
M.E (Kcal/kg)	3000	3000	3000	3000	2900	2900	2900	2900
Methionine	0.48	0.46	0.48	0.44	0.45	0.44	0.43	0.43
Lysine	1.37	1.37	1.38	1.32	1.26	1.23	1.21	1.20
Fibre	2.86	3.50	3.26	3.59	3.09	3.25	3.49	3.70

*Premix contains the following per kilogram of diet: Fe100mg, Mn110mg, Cu 20mg, Zn100mg Se 0.2mg, Menadione 1.33mg, Co 0.6mg, Seno0.6mg, Retinal2000mg, Cholecalciferol2525mg, Atocopherol 25mg, Cobalamin 0.03mg, thiamin 0.83mg, Riboflavin 2 mg, Folic acid 0.33mg, Biotin 0.03, Pantothenic acid 3.75mg, Macin 23.3mg, pydoxine 1.33mg*

### **3.3 Experimental Birds and Management**

Two hundred (200) Ross 308-day-old chicks (unsexed) were raised in an open-sided deep litter housing system. The housing system was bedded with fresh softwood shavings (5 cm deep), and the chicks had *ad libitum* access to starter, grower, and finisher diets as well as water throughout the fifty-six-day experimental period. Upon arrival from the hatchery, chicks of comparable sizes were balanced by weight and randomly distributed among twenty floor pens (each measuring 2.4 m<sup>2</sup>), with five replicate pens per treatment, accommodating ten birds each. Each pen was equipped with plastic feeding and watering troughs. Birds were introduced to twenty-four hours light during the brooding stage through to the Sixth week and then reduced to twelve hours light and twelve hours darkness till the eighth week of the study.

### **3.4 Data Collection**

Data was collected on: growth performance, gut pH carcass characteristics, bone traits and economics of production.

#### **3.4.1 Growth performance**

Two week calculations were made for feed consumption, body weight, body weight gain, feed conversion ratio (feed: gain), and livability. Feed fed was subtracted from leftover to determine feed consumption. By dividing the total pen weight by the total number of birds in the pen, the body weight was calculated. The difference between the birds' final weight and their starting weight was used to compute gain. The FCR was computed by dividing amount consumed during the same period by weight gain. The number of birds presently in each pen was divided by the initial number of birds multiplied by 100 to determine

livability. Throughout the study period, postmortem examinations were performed on deceased birds and the pens were checked twice a day for mortality. In order to account for mortality, feed intake and feed conversion ratio (feed intake divided by weight gain) were calculated.

### **3.4.2 Relative organ weight**

The liver, breast meat, thigh, heart, and empty gizzard weight were taken from the sampled birds and weighed. Each part was expressed as percentage of live body weight (BW).

### **3.4.3 Bone traits**

On days fifty-six post-hatch, the femur and tibiae bones were removed from the left leg of the sampled birds and used to assess bone weight, dimensions, and bone-breaking strength. The dimensions of the femur and tibia were measured, including length (mm) from the proximal end to the distal end and width (mm) at the medial region, using a digital caliper, and the femur and tibia bones of the two sampled birds were weighed (grams) after air drying for one week using an electronic scale (Mettler Toledo) and the result was expressed as a percentage of the live body weight. To determine breaking strength (BS), the flesh was removed manually from the femur and tibiae using a scalpel. Subsequently, the femur and tibiae underwent testing on a universal texture analyzer (Inspekt table50-1, Hegewald and Peschke, Meß- Germany) equipped with a 50 KN load cell and 3-point fixture bed, operating at a test speed of 10 points of data per second. The equipment was controlled by a BenQ computer (24 Inch IPS monitor) using Blue Hill 3 software.

## **3.5 Proximate Analysis of Experimental Diets**

### **3.5.1 Crude protein determination**

The crude protein content of the diets was determined using the AOAC (1990) procedure with a little modification. Protein determination was done in three stages thus digestion, distillation and titration.

#### **Digestion stage**

Two (2.0) grams of diet samples were put into digestion flasks and a half tablet of Selenium-based catalyst was added. 25 ml of concentrated Sulphuric acid was added to each of the flasks and shaken thoroughly to ensure complete soaking of the samples. The flasks were then placed on a digestion burner and heated slowly with intermittent shaking until a clear greenish solution was obtained (usually 1-hour period). The digested samples (digests) were allowed to cool and transferred into a 100 ml volumetric flask and made up to the mark with distilled water.

#### **Distillation**

Kjeldahl nitrogen distillation apparatus (Tecator™ Kjeltex System) was flushed out using distilled water for about 5 minutes. 25 ml of 4 % boric acid was measured into 250 ml conical flasks and two drops of mixed indicator were added. Each of the 100ml diluted digests was transferred into a Kjeldahl distillation tube and then 50ml of 40 % NaOH was added. The apparatus was switched into operation whereby steam was generated and the mixture was heated in a tube. This process liberated gaseous ammonia which was collected into the conical flask until 150ml of the distillate was obtained. The colour change observed

in the conical flask was pink to bluish-green. A blank test was conducted without the test samples.

### **Titration**

The distillates were titrated with 0.1N HCl solution. The end-point was noted during the titration which was from the bluish-green colour to colourless. The volume of HCl used (titre values) was then recorded after colour changes to pink by a drop.

Calculation;

$$\text{Nitrogen (N)} = \frac{(1.4007 \times 0.1 \times \text{Titre value})}{2} \times 10 \quad \text{Crude protein} = \text{N} \times 6.25$$

### **3.5.2 Ash Determination**

Two grams (2g) of diet samples were weighed into already cleaned and weighed crucibles in duplicate. The crucibles were placed in a furnace (Vecstar) preheated at 600°C for two hours (2hrs). The crucibles were removed, cooled, and reweighed. The masses of the crucibles and their contents were found by subtraction. The ash content was calculated by difference and expressed as a percentage of the initial weight of the sample.

### **3.5.3 Crude Fat Determination**

2 g of diet samples were weighed and transferred into a paper thimble plugged at the opening with glass wool and placed into a thimble holder. Petroleum ether of volume 150ml was measured into a previously dried and weighed 250ml round-bottom flask and this was assembled with the thimble holder and its contents. The Quick fit condenser was connected to the Soxhlet Extractor and refluxed for sixteen hours (16hr) on low heat on a heating

mantle. After extraction the flask containing the fat was dried at 105°C in an oven for 30 minutes, cooled in a desiccator and the weight of the fat collected was determined and expressed as a percentage of crude fat (AOAC, 1990).

### **3.5.4 Crude Fibre Determination**

The samples after the determination of the crude fat were weighed and transferred into a 500 ml Erlenmeyer flask and few chips were added and 200 ml of 1.25 % sulphuric acid was added to each of the samples. The flask was then set on a hot plate and connected to a condenser. The content was timed at the onset of boiling. At the end of the thirty minutes, the flask was removed and the contents filtered through a linen cloth in a funnel. Boiling water was used to wash the content till the acid was removed. The distillate was discarded and the residue was placed back into the Erlenmeyer flask containing the chips using a spatula 200 ml of 1.25 % NaOH was added and flask reconnected to the condenser. The content was left to boil for thirty minutes after which the flask was disconnected and the content filtered using linen cloth. The content was washed with boiling water till the base was completely removed and then finally washed with ethanol to break any lumps. The residue was transferred into a porcelain crucible and dried in an oven for 2s hours at 130°C. The weight of the dried residue now composed of the crude fibre and minerals was recorded and then placed in the furnace for 30 minutes to burn off the organic material (crude fibre) leaving the inorganic (minerals). The content after ashing was allowed to cool after which the weight was taken. The crude fibre was obtained by a subtraction of the weight of the ash from the weight of the dried residue before ashing.

### **3.5.5 Metabolisable Energy Determination**

Metabolisable energy (ME kcal/kg) was calculated according to the formula derived by Ponzenga(1985),  $ME \text{ kcal/kg} = (37 * \%CP) + (81.8 * \%EE) + 35 * NFE$  . Where

ME=Metabolisable Energy, CP=Crude protein, EE= Ether Extract and NFE= Nitrogen Free Extract

### **3.5.6 Gastrointestinal pH**

As described by Zanu *et al.* (2023), two birds per replicate pen were sampled and through cervical head dislocation the sampled birds were killed. The birds will be dissected, and the pH of the crop, proventriculus, gizzard, duodenum, jejunum, ileum, and caeca will be measured using a pH tester (Hanna Instruments, UK) by directly inserting the pointed tip into the digesta in the lumen of the proximal end of each segment of the sampled bird, making sure the pH electrode does not touch the walls. The probe were cleaned with distilled water once all of the readings for each bird had been obtained. The next step was calculating the mean of the two readings for each tract section of the two sampled birds.

### **3.5.7 Bone Breaking Strength**

To determine breaking strength (BS), the flesh was removed manually from the femur and tibiae using a scalpel. Subsequently, the femur and tibia will undergo testing on a universal texture analyzer (Inspekt table50-1, Hegewald and Peschke, Meß- Germany) equipped with a 50 KN load cell and 3-point fixture bed, operating at a test speed of 10 points of data per second. The equipment was controlled by a BenQ computer (24 Inch IPS monitor) using Blue Hill 3 software.

### **3.5.8 Cost benefit analysis of birds**

The cost of feed per kg for each of the experimental diets was determined based on the usual market prices of the ingredients at the time of the experiment. The cost of the conventional feed in the diet was determined using the price per kilogram of conventional feed, cost of feed per replicate, cost per kilogram body weight gain, selling price per kilogram of chicken, selling price of broilers and profit on feed. The price per kilogram cost of feed will be determined by multiplying the quantity of each ingredient by the unit cost of the ingredients to get the cost of ingredients. The cost of ingredients was summed up to obtain the total feed cost. Then, the total feed cost was divided by one hundred to obtain the per kilogram cost of feed. The cost of feed per replicate was calculated by multiplying per kilogram cost of feed by the total feed consumed per replicate. The cost per kilogram body weight gain was determined by dividing the cost per bird by kilogram weight gain.

The current selling price per kilogram of chicken was used. The selling price of a bird was determined by multiplying the per kilogram selling price of broilers by the kilogram body weight gain. The profit on feed was calculated by subtracting the cost of feed from the selling price of a bird. The data was subjected to analysis.

### **3.5.9 Organ weight**

The heart, liver, empty gizzard, breast meat and thigh was taken from the sampled birds used for the determination of gut pH and weighed. Each part will be expressed as a percentage of live.

### **3.6 Statistical Analysis of Data**

To evaluate the effect of partial replacement of maize with soya bean oil or full fat soya bean on Ross 308 broiler chicken data were analyzed as a completely randomized design (CRD) using the General Linear Model (GLM) procedure of the Minitab 20.3 statistical software. Tukey's means separation test was used to make comparisons between treatment means at probability  $P < 0.05$ .

## CHAPTER FOUR: RESULTS

### 4.1 Nutrient Composition of Full-Fat Soya Bean Meal

The results of the proximate analysis of the full-fat soya bean meal (FFSBM) gave the following components; 5.35 %, 9.63 %, 7.34 %, 34.15 %, 9.83 %, 7.90 % and 33.14 % for, moisture, ash, crude fat, crude protein, crude fibre and nitrogen free extract respectively (Table 4.1). The test sample's metabolisable energy was 3227.54 Kcal/kg.

**Table 4. 1: Nutrient composition of full-fat soya bean meal**

Nutrient Composition	%
Moisture content	5.35
Ash content	9.63
Crude protein	34.15
Crude fat	9.83
Crude fibre	7.90
Nitrogen free extract	33.14
Metabolisable energy (kcal/kg)	3227.54

### 4.2 Nutrient Composition of Experimental Diets

The proximate analysis of the experimental diets, as presented in Table 4.2, revealed notable variations in nutrient composition across both the starter and grower/finisher phases. Diets formulated with full-fat soya bean (T2) and soya bean oil (T3) generally exhibited higher crude protein levels compared to the control (T1), particularly during the starter phase. Crude fat content was highest in the diet combining conventional and full-fat soya bean (T4) across both feeding phases. However, soya bean oil-based diets (T3) recorded relatively lower metabolisable energy values, especially in the starter phase.

Moisture content was highest in T3 during both feeding phases. Ash content, representing mineral composition, was notably high in T3 diets but declined in T4 during the grower phase, while crude fibre content was highest in T3 and T1 diets. Nitrogen-free extract (NFE), was most abundant in the control diet (T1). Overall, the T4 diet consistently provided the highest metabolisable energy while T3 diets, despite oil inclusion, were relatively less energy-efficient and higher in moisture.

**Table 4. 2: Nutrient Composition of Experimental Diets**

Analyzed Nutrient Composition (%)	Starter Diets				Grower/Finisher Diets			
	T1	T2	T3	T4	T1	T2	T3	T4
Moisture content	8.30	8.18	9.02	8.06	8.01	8.39	10.17	9.81
Ash content	<b>17.25</b>	<b>14.64</b>	<b>19.45</b>	<b>16.85</b>	<b>17.52</b>	<b>17.80</b>	<b>21.49</b>	<b>13.53</b>
Crude protein	<b>17.51</b>	<b>20.58</b>	<b>20.14</b>	<b>19.26</b>	<b>18.83</b>	<b>19.26</b>	<b>19.70</b>	<b>19.48</b>
Crude fat	9.32	8.76	8.29	10.09	9.53	7.14	10.96	11.14
Crude fibre	4.79	7.72	8.57	5.86	<b>9.38</b>	<b>8.18</b>	<b>6.67</b>	<b>6.37</b>
Nitrogen free extract	42.83	40.12	34.53	39.88	36.73	39.23	31.01	39.67
Metabolisable energy kcal/kg	2909	2882	<b>2632</b>	2934	2762	2670	2711	3021

*T1= control diet (conventional soya bean), T2= Full-fat soya bean diet, T3 = Soya bean oil diet, and T4 = Conventional soya bean + full fat soya bean diet.*

*Metabolisable energy (MEkcal/kg) was calculated according to the formula derived by Pauzenga (1985: MEkcal/kg= (37\*%CP)+(81.8\*%EE)+35\*NFE)*

*Where: CP=crude protein, CF=crude fibre, EE=ether extract, NFE= nitrogen free extract, DM=dry matter, MC =moisture content*

### 4.3 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean oil on the Growth Performance of Broiler Chickens at d 14

The results in Table 4.3 show that there were no statistically significant differences ( $P > 0.05$ ) among the treatment groups for all measured parameters, including body weight (BW), body weight gain (BWG), feed intake, feed conversion ratio (FCR), and livability during the starter phase (day 0–14). Although numerical differences were observed such as the highest BW (201.4 g) and feed intake (283.8 g) in T3 (soya bean oil diet), and the lowest (efficient) FCR (1.900) in T2 (full-fat soya bean diet) these variations were not significant. Livability remained high across all treatments, ranging from 98 % to 100 %.

**Table 4. 3: Effect of Partial Replacement of Maize with Full Fat Soya bean or Soya bean Oil on the Growth Performance of Ross 308 Broiler Chicken at D 0-14**

Treatment	BW(g)	BW Gain(g)	Feed Intake(g)	FCR (g)	Livability (%)
T1	146.8	158.0	256.0	1.977	100.0
T2	187.0	148.0	235.0	1.900	100.0
T3	201.4	163.4	283.8	2.053	98.00
T4	188.1	152.1	251.3	1.950	98.00
<b>SEM</b>	24.80	18.30	23.80	0.114	1.410
<b>P-value</b>	0.459	0.937	0.552	0.817	0.585

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, BW=body weight, and FCRc= corrected feed conversion ratio.

#### 4.4 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean oil on the Growth Performance of Ross 308 Broiler

##### Chickens at d 28

According to results in Table 4.4, feed intake differed significantly ( $P = 0.003$ ) among the treatment groups, with T2 (full-fat soya bean diet) recording the lowest intake (938.0 g), while T1 (control), T3 (soya bean oil diet), and T4 (conventional + full-fat soya bean diet) had significantly higher and similar feed intake values. No significant differences ( $P > 0.05$ ) were observed in body weight, body weight gain, feed conversion ratio (FCR), or livability across treatments. Body weight ranged from 534.0 g (T2) to 641.0 g (T1), and FCR ranged from 1.966 (T2) to 2.375 (T3). Livability was high across all treatments, ranging from 98 % to 100 %.

**Table 4. 4: Effect of partial replacement of maize with full fat soya bean or soya bean oil on the growth performance of Ross 308 broiler chicken at d 0-28**

Treatment	BW(g)	BW Gain(g)	Feed Intake(g)	FCR(g)	Livability (%)
T1	641.0	570.0	1303.0 <sup>a</sup>	2.304	100.0
T2	534.0	495.0	938.0 <sup>b</sup>	1.966	100.0
T3	615.0	577.0	1366.9 <sup>a</sup>	2.375	98.00
T4	637.2	601.2	1303.0 <sup>a</sup>	2.175	98.00
<b>SEM</b>	33.60	40.70	74.40	0.136	1.410
<b>P –value</b>	0.127	0.318	0.003	0.196	0.585

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, BW=body weight, and FCRc= corrected feed conversion ratio.

#### 4.5 Effect of Partial Replacement of Maize with Full Fat Soya beans or Soya Bean Oil on the Growth Performance of Ross 308 Broiler Chickens at d 42

The results in Table 4.5 show that at day 42, there were significant differences ( $P < 0.05$ ) among the treatment groups for body weight (BW), body weight gain (BWG), and feed intake. T2 (full-fat soya bean diet) recorded the lowest values for BW (763.0 g), BWG (724.0 g), and feed intake (1887 g), all of which were significantly lower than those observed in T1 (control), T3 (soya bean oil diet), and T4 (conventional + full-fat soya bean diet). However, no significant differences ( $P > 0.05$ ) were found for feed conversion ratio (FCR) and livability among the treatments. FCR values ranged from 2.391 (T3) to 2.723 (T4), while livability remained high across treatments, ranging from 98 % to 100 %.

**Table 4. 5: Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya bean Oil on the Growth Performance of Ross 308 Broiler Chicken at D 42**

Treatment	BW(g)	BW Gain(g)	Feed Intake(g)	FCR(g)	Livability (%)
T1	1141 <sup>a</sup>	1105 <sup>a</sup>	2641 <sup>a</sup>	2.399	100.0
T2	763.0 <sup>b</sup>	724.0 <sup>b</sup>	1887 <sup>b</sup>	2.643	100.0
T3	1069 <sup>a</sup>	1031 <sup>a</sup>	2443 <sup>ab</sup>	2.391	98.00
T4	957.8 <sup>a</sup>	921.8 <sup>a</sup>	2508 <sup>ab</sup>	2.723	98.00
SEM	47.50	47.60	155	0.194	1.410
P-VAL	0.000	0.000	0.017	0.533	0.585

<sup>abc</sup>Means with different super scripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, BW=body weight, and FCRc= corrected feed conversion ratio.

## 4.6 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean oil on the Growth Performance of Ross 308 Broiler Chickens at d 49

At day 49, broilers fed the control diet (T1) showed significantly ( $P < 0.05$ ) higher body weight and weight gain compared to those fed full fat soya bean diet(T2), which recorded the lowest values for both parameters, while birds on the soy bean oil diet (T3) and the conventional soya bean + full-fat soya bean combination diet (T4) had intermediate values. Feed intake was significantly ( $P < 0.05$ ) higher in T1 and T3 compared to T2, with T4 not differing significantly from either. Feed conversion ratio (FCR) and livability were not significantly affected by the dietary treatments ( $P > 0.05$ ). (Table 4.6)

**Table 4. 6: Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Growth Performance of Ross 308 Broiler Chicken at D 49**

Treatment	BW(g)	BW Gain(g)	Feed Intake(g)	FCR(g)	Livability (%)
T1	1387 <sup>a</sup>	1351 <sup>a</sup>	3346 <sup>a</sup>	2.479	100.0
T2	946 <sup>c</sup>	907 <sup>c</sup>	2562 <sup>b</sup>	2.831	100.0
T3	1278 <sup>ab</sup>	1240 <sup>ab</sup>	3178 <sup>ab</sup>	2.563	98.00
T4	1189 <sup>b</sup>	1153 <sup>b</sup>	3195 <sup>ab</sup>	2.784	98.00
<b>SEM</b>	32.10	32.10	160.0	0.161	1.410
<b>P-value</b>	0.000	0.000	0.015	0.372	0.585

*Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, BW=body weight, and FCRc= corrected feed conversion ratio.*

#### 4.7 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Growth Performance of Ross 308 Broiler Chickens at d 56

At d 56, there were significant differences ( $P < 0.05$ ) among treatments for body weight (BW), body weight gain (BWG), feed intake, and corrected feed conversion ratio (FCR), while livability remained unaffected ( $P > 0.05$ ). T2 (full-fat soya bean diet) recorded significantly lower BW (1178 g), BWG (1139 g), and feed intake (3798 g) compared to the other treatments. T1 (control), T3 (soya bean oil diet), and T4 (conventional + full-fat soya bean diet) showed significantly higher BW and BWG, with T1 and T4 also showing the highest feed intake. FCR was highest in T2 (3.338), and lowest in T1 (2.582). Livability remained high across all treatments, ranging from 98 % to 100 % (Table 4.7).

**Table 4. 7: Effect of Partial Replacement of Maize with Full Fat Soya beans or Soya Bean Oil on the Growth Performance of Ross 308 Broiler Chicken at D 56**

Treatment	BW(g)	BW Gain(g)	Feed Intake(g)	FCRc	Livability (%)
T1	1699 <sup>a</sup>	1663 <sup>a</sup>	4611 <sup>a</sup>	2.582 <sup>b</sup>	100.0
T2	1178 <sup>b</sup>	1139 <sup>b</sup>	3798 <sup>b</sup>	3.338 <sup>a</sup>	100.0
T3	1519 <sup>a</sup>	1481 <sup>a</sup>	4330 <sup>ab</sup>	2.637 <sup>ab</sup>	98.0
T4	1582 <sup>a</sup>	1546 <sup>a</sup>	4589 <sup>a</sup>	2.970 <sup>ab</sup>	98.00
<b>SEM</b>	56.40	56.5	185	0.128	1.410
<b>P-value</b>	0.000	0.000	0.0230	0.045	0.585

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, BW=body weight, and FCRc= corrected feed conversion ratio.

#### **4.8 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Gut pH of Ross 308 Broiler Chickens**

The effect of partial replacement of maize with full-fat soya beans and soya bean oil on gut pH in broiler chickens revealed no statistically significant differences ( $P > 0.05$ ) across all gut sections as shown in Table 4.8. In the crop, the highest pH was observed in birds fed the soya bean oil diet (T3 = 4.902), while the lowest was in birds fed the full-fat soya bean diet (T2 = 4.529). In the proventriculus, birds fed the combination of conventional and full-fat soya bean (T4) had the highest pH (1.517), and those on the control diet (T1) had the lowest (1.227). For the gizzard, pH ranged from 1.600 (T4) to 1.840 (T1), with the control group having the highest value. In the duodenum, pH values were very similar across treatments, ranging from 4.552 (T2) to 4.703 (T4). In the jejunum, the control diet (T1) again showed the highest pH (5.694), and soya bean oil group (T3) the lowest (5.595). In the ileum, the most notable numerical difference was observed. Birds on the full-fat soya bean diet (T2) had the highest pH (6.420), while those on the control diet (T1) had the lowest (5.893). Although not statistically significant ( $P = 0.189$ ), this is the largest observed numerical shift. Lastly, in the caeca, pH ranged from 6.735 (T1) to 6.872 (T2).

**Table 4. 8: Effect of Partial Replacement of Maize with Full Fat Soya beans or Soya Bean Oil on the Gut pH Of Ross 308 Broiler Chickens**

Treatment	Crop	Proventriculus	Gizzard	Duodenum	Jejunum	Ileum	Caeca
T1	4.80 <sup>1</sup>	1.227	1.840	4.569	5.694	5.893	6.735
T2	4.52 <sup>9</sup>	1.460	1.759	4.552	5.658	6.420	6.872
T3	4.90 <sup>2</sup>	1.454	1.770	4.653	5.595	6.370	6.853
T4	4.62 <sup>3</sup>	1.517	1.600	4.703	5.642	6.377	6.825
<b>SEM</b>	0.17 <sup>9</sup>	0.196	7.910	0.168	0.129	0.186	0.096
<b>P-value</b>	0.46 <sup>7</sup>	0.735	0.416	0.909	0.958	0.189	0.756

<sup>abc</sup> Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), , T1= control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, and T4 = Conventional soya bean + full fat soya bean diet.

#### **4.9 Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Carcass Traits (% BW) of Ross 308 Broiler Chickens**

The partial replacement of maize with full-fat soya beans and soya bean oil in the diets of broiler chickens did not significantly ( $P > 0.05$ ) influence the relative weights of the heart, liver, breast, thigh, and spleen as percentages of body weight (Table 4.9). However, significant differences were observed in the gizzard and fat pad percentages ( $P = 0.014$  and  $P = 0.000$ , respectively). The gizzard weight was highest in birds fed full fat-soya bean diet

(T2), significantly differing from those on the soya bean oil diet (T3), which had the lowest value. For fat pad percentage, birds on the soya bean oil diet (T3) had the highest value, significantly greater than those on the full fat soya bean diet (T2) and the combined soya bean diets (T4), while the control diet (T1) had an intermediate value.

**Table 4. 9: Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Carcass Traits (% BW) of Ross 308 Broiler Chickens**

<b>Treatment</b>	<b>Heart</b>	<b>Liver</b>	<b>Gizzard</b>	<b>Breast</b>	<b>Thigh</b>	<b>Spleen</b>	<b>Fat pad</b>
T1	0.701	2.030	2.003 <sup>sb</sup>	3.759	9.522	0.118	0.011 <sup>ab</sup>
T2	0.824	2.070	2.323 <sup>a</sup>	3.607	9.448	0.010	0.009 <sup>bc</sup>
T3	0.795	4.010	1.897 <sup>b</sup>	3.906	9.660	0.143	0.012 <sup>a</sup>
T4	0.798	2.050	2.232 <sup>ab</sup>	3.967	9.486	0.125	0.007 <sup>c</sup>
<b>SEM</b>	0.032	1.010	0.090	0.199	0.163	0.018	0.001
<b>P-value</b>	0.076	0.444	0.014	0.592	0.808	0.411	0.000

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, and T4 = Conventional soya bean + full fat soya bean diet.

#### **4.10 Effect of Partial Replacement of Maize with Full Fat Soya beans or Soya bean oil on the Bone Traits of Ross 308 Broiler Chickens**

From Table 4.10, there was a significant ( $P < 0.05$ ) effect of the treatment on tibia length, with birds fed the control diet (T1), full-fat soya bean diet (T2), and the combined soya bean diet (T4) exhibiting significantly longer tibiae than those on the soya bean oil diet (T3), while no significant ( $P > 0.05$ ) differences were observed among treatments for femur length, tibia diameter, femur diameter, tibia and femur breaking strength.

**Table 4. 10: Effect of Partial Replacement of Maize with Full Fat Soya Beans or Soya Bean Oil on the Bone Traits of Ross 308 Broiler Chickens on Day 56**

Treatment	Tibia length(mm)	Femur Length(mm)	Tibia Diameter(mm)	Femur Diameter(mm)	Tibia BS	Femu r BS
T1	98.80 <sup>a</sup>	71.62	6.445	7.368	134.3	209.6
T2	98.14 <sup>a</sup>	69.32	6.201	6.793	152.0	196.2
T3	92.14 <sup>b</sup>	72.29	6.583	7.204	188.2	207.7
T4	96.11 <sup>a</sup>	71.28	6.290	7.124	142.9	200.7
<b>SEM</b>	0.898	1.060	0.194	0.201	15.60	18.20
<b>P-Value</b>	0.000	0.267	0.532	0.267	0.115	0.949

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ) T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean diet, and BS=Breaking strength.

#### **4.11 Effect of Partial Replacement of Maize with Full Fat Soya beans or Soya Bean Oil on the Economic Efficiency of Ross 308 Broiler Chickens**

The economic analysis revealed statistically significant differences ( $P < 0.05$ ) in several economic parameters among the dietary treatments, including Price of bird at day 56, Profit, Feed cost to profit ratio, Price-to-feed cost ratio, and Performance Efficiency Index (PEI), while Total feed cost and cost per kg feed did not differ significantly ( $P > 0.05$ ).

The Price of birds at day 56 was significantly higher ( $P = 0.000$ ) in T1 (C59.47), T3 (C53.15), and T4 (C55.36) compared to T2 (C41.23), which recorded the lowest value. Birds in T1, T3, and T4 had statistically similar price values. Although the Total Feed Cost

did not differ significantly among treatments ( $P = 0.458$ ), numerically, T1 had the highest cost (C42.88), followed by T3 (C40.96), T4 (C39.90), and T2 (C38.96). Profit at day 56 differed significantly ( $P = 0.000$ ), with T1 (C16.59), T3 (C12.20), and T4 (C15.46) yielding significantly higher profits than T2 (C2.27). The Feed cost to profit ratio showed significant differences ( $P = 0.001$ ). T1 (0.3887), T3 (0.2994), and T4 (0.3939) recorded higher ratios compared to T2 (0.0704), which was significantly lower.

Similarly, the Price-to-feed cost ratio was significantly different ( $P = 0.001$ ). T1 (1.3887), T3 (1.2994), and T4 (1.3939) had comparable values and were significantly higher than T2 (1.0704), which was statistically different.

Finally, the Performance Efficiency Index (PEI) varied significantly among treatments ( $P = 0.000$ ). T1 had the highest PEI value (126.1), followed by T3 (94.84) and T4 (93.52), which were statistically similar, while T2 recorded the lowest PEI (67.55) and was significantly different from the other treatment diets.

**Table 4. 11: Effect of Partial Replacement of Maize with Full Fat Soya bean or Soya bean Oil on Economic Efficiency of Broiler Chickens**

Treatment	Cost per kg feed	Price of bird d56 ₪	Total Feed Cost d56 ₪	Profit d56 ₪	Profit d56: Feed cost ₪	Price/Wt, ₪ Feed Cost ₪
T1	7.2631	59.47 <sup>a</sup>	42.88	16.59 <sup>a</sup>	0.3887 <sup>a</sup>	1.3887 <sup>a</sup>
T2	7.2364	41.23 <sup>b</sup>	38.96	2.27 <sup>b</sup>	0.0704 <sup>b</sup>	1.0704 <sup>b</sup>
T3	7.1560	53.15 <sup>a</sup>	40.96	12.20 <sup>a</sup>	0.2994 <sup>a</sup>	1.2994 <sup>a</sup>
T4	7.2631	55.36 <sup>a</sup>	39.90	15.46 <sup>a</sup>	0.3939 <sup>a</sup>	1.3939 <sup>a</sup>
<b>SEM</b>	0.0414	1.970	1.760	1.890	0.051	0.051
<b>P-value</b>	0.135	0.000	0.458	0.000	0.001	0.001

<sup>abc</sup>Means with different superscripts on the same column are Significantly different ( $P < 0.05$ ), T1 = control diet (conventional soya bean), T2 = Full-fat soya bean diet, T3 = Soya bean oil diet, T4 = Conventional soya bean + full fat soya bean die.

## **CHAPTER FIVE: DISCUSSION**

### **5.1 Nutrient Composition of Full –Fat Soya Bean**

Analysis of the full –fat soya bean (FFSB) used in this study revealed 34.15% crude protein 9.83% crude fibre, 7.34% crude fat, and 33.14% nitrogen –free extract (NFE), with a calculated metabolisable energy (ME) of 3144kcal/kg, confirming its value as an energy-rich feed ingredient. The crude protein level indicates that FFB is a reliable protein source. Km et al. (2021) reported that crude protein value ranging from 33% to 38% in full –fat soya bean, supporting their potential as alternative to soya bean meal when properly heat-processed to deactivate anti-nutritional factors like trypsin inhibitors. Similarly, Oduguwa et al. (2014) found comparable protein content in heat-treated full-fat soya bean, further validating its implication in poultry diet. The crude fat level of 7.3% is noteworthy, as it plays a vital role in increasing the energy concentration of the diet.

Dietary fat not only boosts caloric supply but also facilitates the absorption of fat-soluble vitamins while minimizing feed dustiness (Sibbald, 1986). Although this value is slightly lower than the 10-18% ether extract range reported by Olukosi et al. (2008) for full-fat soya bean under different processing methods, it still contributed positively to the metabolisable energy (ME), which was calculated at 3144kcal/kg. This figure is below the NRC (1994) value estimates of 3200-3400 kcal/kg for full-fat soya bean, suggesting that the sample analysed was effectively processed and preserved much of its inherent oil content. The nitrogen-free extract (33.14%) and crude fibre (9.83%) values illustrate the carbohydrate profile of the FFSB. The fibre level, though higher than that of conventional soya bean meal (<5% CF), is typical of full-fat forms due to hull retention. Elevated fibre can reduce nutrient

digestibility, but appropriate inclusion levels help maintain a balance between energy and fibre (Adeyemi and Ajayi, 2020). The NFE component, composed of soluble carbohydrates and starches, contribution readily available energy to support rapid broiler growth (Rostagno et al., 2017). Moisture (5.35%) and ash (9.63%) content were within acceptable feed standard; low moisture promotes safe storage by limiting microbial growth, while the moderately high ash may result from mineral –rich soil or hull retention. Since excessive ash can reduce dietary nutrient density, its level should be carefully managed (Kellem and church, 2010).

## **5.2 Nutrient Composition of the Experimental Diet**

The proximate composition of the experimental diet indicated that though the analyzed crude protein and crude fibre were slightly lower than the formulated level, the diet were similar.

## **5.3 Effect of Partial Replacement of Maize with Full –Fat Soya Bean or Soya Oil on Growth Performance of Broiler 308 Broiler Chickens at d14**

The study showed that partial replacement of maize with full –fat soya bean (FFSB) or soya oil influence growth performance differently across production stages during the starter phase (d0-d14), no significant differences were found in body weight, body weight gain, feed intake, feed conversion ratio, or livability, although numerical variation indicated trend. Birds on soya bean oil diet recorded the highest BW and feed intake, while those on FFSB diet had the lowest FCR, consistent with Batal and Parson (2004), who highlighted improve early nutrient digestibility with oil inclusion.

On day 28, only feed intake differed significantly with FFBSB group consuming less, likely due to higher diet energy density causing early satiety (Leeson and Summers, 2001). Despite reduced intake, BW and BWG were unaffected, suggesting efficient nutrient utilization.

At d42, birds on FFBSB shown lower BW, BWG and feed intake, suggesting cumulative effects of reduced palatability or anti-nutritional factor (Sundu et al., 2006). By d49, performance differences widened, with the control group outperforming soya bean oil diet in BW and BWG, possibly due to poor oil utilization or nutrient imbalance (Zhang et al., 2011). FFBSB and mixed diet performed moderately.

At d56, significant depression in BW, BWG, FI and FCR in the FFBSB group reinforced its long-term nutritional in- balance and anti- nutritional impact (OlomU., 2011). Livability remained high (98 -100%) across treatment indicating diet safety.

#### **5.4 Effect of Partial Replacement of Maize with Full Fat Soya Bean or Soya Oil on Gut pH**

Partial replacement of maize with FFBSB or soya oil did not significantly alter gut pH, though numerical variation occurred in certain segment. in the crop, the highest pH was recorded in FFBSB-fed birds, while soya bean oil diet yielded the lowest, consistent with Syihus (2011), who reported that fat-induced alterations in fermentation and gastric emptying. proventriculus and gizzard pH values remain acidic (1.2-1.8), reflecting normal gastric function, with slight diet-related variation likely due to protein and fat buffering (Rayindran et al., 2006)

Small intestine pH remain moderately alkaline, consistent with neutralization by pancreatic and bile secretion. Only minor difference were observed in the duodenum and jejunum, while ileal pH was highest in soya bean oil fed birds, suggesting reduce fermentation or bile salt-related effect (Hetland et al.,2004). Caecal pH remained stable across treatments (6.7-6.9) typical of high gut fermentation and consistent with Torok et al. (2011), indicating minimal impact of dietary fat on microbial activity.

## **5.5 Effects of Partial Replacement of Maize with Full Fat Soya Bean or Soya Oil on Carcass Traits**

Maize replacement with FFSB or soya bean oil did not significantly affect relative weight of most carcass components (heart, liver, breast, thigh, and spleen). These results align with Fan et al. (2008) and Rayindran et al. (2016), who found out that soya bean –based ingredient support growth without altering organ development when diet is isoenergetic and isonitrogenous.

However, significant differences were observed in gizzard and fat weight. Birds on soya bean oil diet showed large fat pad due poor utilization of energy. This is in agreement with a report made by Bello et al. (2023) who found out that excess energy is converted to fat after maintenance and production. FFSB-fed birds showed the largest gizzard weight likely due to had dietary fibre availability compared to the soya bean oil diet (Zhao et al., 2009, Zhang et al., 2011) intermediate value in control and mixed diet reinforced the role of form and availability in lipid deposition (Zeng et al., 2015).

These indigestible fractions stimulate gizzard activity and muscular development, enhancing its relative size compared to diets containing extracted oils. The fibrous matrix and non-starch polysaccharides present in full-fat soya bean exert a mechanical effect on the gastrointestinal tract, thereby promoting hypertrophy of the gizzard (Ravindran, 2013; Choct, 2018).

## **5.6 Effects of Partial Replacement of Maize with Full Fat Soya Bean or Soya Oil on Bone Traits**

The diets significantly influenced tibia length but not femur length, bone diameter or breaking strength, birds on the control, FF SB, and mixed diet had longer tibia compared to those on soya bean oil diet the diet are balance in protein – energy profile alongside potential bioactive benefit for mineralization (Wei et al., 2013). Reduced tibia length in the soya bean oil group may reflect imbalances in protein – energy ratio or mineral metabolism linked to rapid energy released from free oils (Fritts and Waldroup, 2003; Korver, 2005). The reduced tibia length in T3 is primary due its effect on nutrient partitioning and bone mineralization. This is in compliance with Rafeeq *et al.*, 2020). et al. (2020) who reported that dietary oil increase the density of feed, it may alter calcium and phosphorus utilization, which are essential for proper skeletal development. Excess dietary fat can also interfere with some vitamin metabolism, thereby reduce mineral absorption efficiency and ultimately compromising tibia growth and mineral deposition (Olukosi et al., 2019) reported.

## **5.7 Effects of Partial Replacement of Maize with Full Fat Soya Bean or Soya Oil on Economic Efficiency**

Economic outcomes were strongly influenced by diet. Birds on conventional soya bean (T1) soya bean oil (T3), and combined diet (T4) achieved higher market price and profit compare to those on FF SB diet (T2). This agrees with Tijani et al. (2016) and Oladokun et al. (2018), who linked improved growth and carcass traits to higher economic returns. Feed cost were comparable across treatments, indicating maize substitution with soya bean derivative did not raise cost, echoing Ojebiyi et al. (2014). Profit and performance efficiency index (PEI) were highest in T1, followed by T3 and T4, reflecting better growth and feed efficiency (Attia et al., 215). T2 's lower profit highlighted that feed cost alone is less important than market weight in determining returns (Onu and Madubuike, 2012). Overall, soya bean oil and combined diet improved cost efficiency, while sole FF SB use was less profitable due to performance limitation, reinforcing the need for balanced formulation (Adeniji and Jimoh, 2007).

## **CHAPTER SIX: CONCLUSION AND RECOMMENDATIONS**

### **6.1 Conclusions**

From the results of the 8-week trial, the following conclusions were made:

Partial replacement of maize with full –fat soya bean (FFSB) or soya bean oil (SO) has varied effects on broiler growth across the production cycle. While performance was similar across treatment during the starter phase (day 14), differences emerged from day 28 onward. Birds fed FFSB consistently exhibited lower feed intake, body weight, and body weight gain, especially by day 42, 49, and 56, showing reduced growth with advancing age. In contrast, the control diet yielded the best growth while soya beans oil showed intermediate performance.

Gut pH values were not significantly affected, suggesting that substituting maize with soya bean products does not disturb gastrointestinal pH balance.

Carcass traits were influenced, FFSB diet increasing gizzard weight whereas soya bean oil diet led to higher abdominal fat deposition.

Bone traits showed that only tibia length was affected, being shorter in birds on soya bean oil diet. Femur length, bone diameters, and breaking strength remained unaffected.

Soya bean oil diet and mixed diet (regular soya bean and full-fat soya bean) diets resulted in increased price per bird, profit, and performance efficiency index (PEI) compared to birds on full-fat soya bean diet.

## **6.2 Recommendations**

Based on the findings, the following recommendations are made

1. For optimal broiler performance, particularly at the starter phase, partial inclusion or blending with conventional soya beans meal is recommended.
2. Soya bean oil should be incorporated carefully, as it may increase fat deposition while reducing tibia length.
3. Using balanced blend FFSB, soya bean oil, and conventional soya bean by reducing maize inclusion in poultry diet is advisable to optimize growth, carcass quality, skeletal development, and fat deposition in broilers.

## REFERENCES

- Aardsma, M. P., Mitchell, R. D., and Parsons, C. M. (2017). Relative metabolisable energy values for fats and oils in young broilers and adult roosters. *Poultry Science*, *96*(7), 2320-2329.
- Ababor, S., Tamiru, M., Alkhtib, A., Wamatu, J., Kuyu, C. G., Tekka, T. A., ... & Burton, E. (2023). The use of biologically converted agricultural by-products in chicken nutrition. *Sustainability*, *15*(19), 14562.
- Abbas, T. E. E., Elamin, K. M., and Rahmatalla, S. A. (2019). Effect of replacing maize with full-fat soya bean on broiler performance and carcass yield. *Sudan Journal of Animal Production*, *18*(1), 33–41.
- Abdel-Raheem, S. M., Mohammed, E. S. Y., Mahmoud, R. E., El Gamal, M. F., Nada, H. S., El-Ghareeb, W. R., ... and Kishawy, A. T. (2023). Double-fermented soybean meal totally replaces soybean meal in broiler rations with favorable impact on performance, digestibility, amino acids transporters and meat nutritional value. *Animals*, *13*(6), 1030.
- Abdel-Raheem, S. M., Mohammed, E. S. Y., Mahmoud, R. E., El Gamal, M. F., Nada, H. S., El-Ghareeb, W. R., ... & Kishawy, A. T. (2023). Double-fermented soybean meal totally replaces soybean meal in broiler rations with favorable impact on performance, digestibility, amino acids transporters and meat nutritional value. *Animals*, *13*(6), 1030.
- Abolhasani, M., Rahimi, G., and Gholami, M. (2020). Effects of solvent-extracted soya bean meal on broiler growth performance and carcass yield. *Poultry Science*, *99*(7), 3211-3219.

- Adebowale, T. O., Yao, K., and Oso, A. O. (2019). Major cereal carbohydrates in relation to intestinal health of monogastric animals: *A review. Animal nutrition, 5(4), 331-339.*
- Adeniji, A. A., and Jimoh, B. (2007). Effects of replacing maize with enzyme-supplemented cassava root sieviate meal on the performance of broiler chickens. *Pakistan Journal of Nutrition, 6(3), 256–260.*
- Adeola, O., and Cowieson, A. J. (2016). Board-invited review: Opportunities and challenges in using exogenous enzymes to improve nonruminant animal production. *Journal of Animal Science, 94(1), 1–21.*
- Adeola, O., and Cowieson, A. J. (2021). Opportunities and challenges in using exogenous enzymes to improve amino acid availability in animal feeds. *Animal Feed Science and Technology, 271, 114705.*
- Adewole, D. I., Rogiewicz, A., Dyck, B., and Slominski, B. A. (2016). Chemical and nutritive characteristics of canola meal from Canadian processing facilities. *Animal Feed Science and Technology, 222, 17–30.*
- Adeyemi, O. A., and Ajayi, O. O. (2020). Nutritional value and effects of dietary inclusion of full-fat soya bean in broiler chicken diets: A review. *Nigerian Journal of Animal Science, 22(3), 1–13.*
- Adeyemo, G. O., and Longe, O. G. (2018). Effects of soya bean products on performance and economic returns in broiler chickens. *International Journal of Poultry Science, 17(3), 101–108*

- Adeyemo, G. O., Ige, A. O., and Afolabi, K. D. (2020). Performance characteristics of broilers fed raw and heat-treated full-fat soya beans. *Nigerian Journal of Animal Science*, 22(3), 154-163.
- Affonfere, M., Chadare, F. J., Fassinou, F. T. K., Linnemann, A. R., and Duodu, K. G. (2023). In-vitro digestibility methods and factors affecting minerals bioavailability: a review. *Food Reviews International*, 39(2), 1014-1042.
- Afolayan, S. B., Dafwang, I. I., Tegbe, T. S., and Sekoni, A. (2015). Response of broiler chickens fed on maize-based diets substitute.pdf. *Asian Journal of Poultry Science*, 6(1), 15–22.
- Aguihe, P. C., and Isikwenu, J. O. (2016). Growth performance and carcass characteristics of broiler finisher birds fed full-fat soya beans and cottonseed meal diets. *Nigerian Journal of Animal Science*, 18(1), 56–62.
- Ahiwe, E. U., Omede, A. A., Abdallah, M. B., & Iji, P. A. (2018). Managing dietary energy intake by broiler chickens to reduce production costs and improve product quality. *Animal husbandry and nutrition*, 115, 115-145.
- Ahmad, T., Khan, M. Z., Rehman, H., and Iqbal, M. (2020). Probiotics and their impact on broiler performance: A review. *Journal of Applied Poultry Research*, 29(3), 813-823.
- Ahmed, H. G. M. D., Naeem, M., Faisal, A., Fatima, N., Tariq, S., & Owais, M. (2023). Enriching the content of proteins and essential amino acids in legumes. In *Legumes biofortification* (pp. 417-447). Cham: Springer International Publishing.

- Akter, M. S., Uddin, M. T., and Dhar, A. R. (2023). Advancing safe broiler farming in Bangladesh: An investigation of management practices, financial profitability, and consumer perceptions. *Commodities*, 2(3), 312-328.
- Al Loman, A., and Ju, L. K. (2016). Soya bean carbohydrate as fermentation feedstock for production of biofuels and value-added chemicals. *Process Biochemistry*, 51(8), 1046-1057.
- Alagawany, M., Elnesr, S. S., and Farag, M. R. (2018). The role of exogenous enzymes in promoting growth and improving nutrient digestibility in poultry. *Iranian journal of veterinary research*, 19(3), 157.
- Alagawany, M., Elnesr, S. S., and Farag, M. R. (2022). The beneficial applications of soya bean and its derivatives in poultry nutrition: An updated review. *Poultry Science*, 101(7), 101996. <https://doi.org/10.1016/j.psj.2022.101996>
- Alagawany, M., Elnesr, S. S., Farag, M. R., and Dawood, M. A. (2019). Nutritional strategies to alleviate heat stress in broilers: A comprehensive review. *World's Poultry Science Journal*, 75(1), 33-50.
- Algarni, T. A., Osman, R. H., Zomrawi, W. B., and Abdalhag, M. A. (2020). Effect of phase feeding on broiler performance. *Online Journal of Animal and Feed Research*, 10(1), 36–40.
- Ali, A., Al-Badwi, H., Al-Sharafi, M. A., and Alqhtani, A. H. (2021). Effects of replacing soya bean oil with poultry fat on broiler performance and carcass traits. *Journal of Animal Physiology and Animal Nutrition*, 105(1), 120–127.

- Ali, M. (2022). Effects of High Oleic Full-Fat Soya bean on Live Performance, Carcass and Meat Quality, Meat Fatty Acid Composition, Amen and Amino Acid Sid in Broilers. *North Carolina State University*.
- Al-Marzooqi, W., and Leeson, S. (2016). Evaluation of different dietary energy levels on growth performance and carcass characteristics of broilers. *International Journal of Poultry Science*, *15(1)*, 1–6.
- Amer, S. A., Beheiry, R. R., Abdel Fattah, D. M., Roushdy, E. M., Hassan, F. A., Ismail, T. A., ... & Metwally, A. E. (2021). Effects of different feeding regimens with protease supplementation on growth, amino acid digestibility, economic efficiency, blood biochemical parameters, and intestinal histology in broiler chickens. *BMC Veterinary Research*, *17(1)*, 283.
- Amerah, A. M., Ravindran, V., and Lentle, R. G. (2007). Feed particle size: Implications on the digestion and performance of poultry. *World's Poultry Science Journal*, *63(3)*, 439–455.
- Amerah, A. M., Van de Belt, K., & van Der Klis, J. D. (2015). Effect of different levels of rapeseed meal and sunflower meal and enzyme combination on the performance, digesta viscosity and carcass traits of broiler chickens fed wheat-based diets. *Animal*, *9(7)*, 1131-1137.
- An, S. H., Kim, D. W., and An, B. K. (2016). Effects of dietary calcium levels on growth performance, bone characteristics, and calcium metabolism in broiler chickens. *Poultry Science*, *95(6)*, 1357–1365. <https://doi.org/10.3382/ps/pew078>

- Andam, K. S., Johnson, M. E., Ragasa, C., Kufoalor, D. S., and Das Gupta, S. (2017). A chicken and maize situation: the poultry feed sector in Ghana (Vol. 1601). *Intl Food Policy Res Inst.*
- Anderson, J. W., Johnstone, B. M., and Cook-Newell, M. E. (1995). Meta-analysis of effect of soy protein intake on serum lipids. *New England of Medicine* 333(5), 276-285.
- Anyiam, P. N., Phongthai, S., Sai-Ut, S., Kingwascharapong, P., Jung, Y. H., Zhang, W., & Rawdkuen, S. (2025). Nutritional Components and Digestibility Profiles of Some Potential Plant-Based Protein Sources. *Foods*, 14(10), 1769.
- Apajalahti, J., and Vienola, K. (2016). Interaction between chicken intestinal microbiota and protein digestion. *Animal Feed Science and Technology*, 221, 323–330. <https://doi.org/10.1016/j.anifeedsci.2016.04.002>
- Arteaga-Wences, Y. J., Estrada-Angulo, A., Ríos-Rincón, F. G., Castro-Pérez, B. I., Mendoza-Cortéz, D. A., Manriquez-Núñez, O. M., ... & Plascencia, A. (2021). The effects of feeding a standardized mixture of essential oils vs monensin on growth performance, dietary energy and carcass characteristics of lambs fed a high-energy finishing diet. *Small Ruminant Research*, 205, 106557.
- Asghar, M. U., Sajid, Q. U. A., Wilk, M., Konkol, D., & Korczyński, M. (2024). Influence of various methods of processing soybeans on protein digestibility and reduction of nitrogen deposits in the natural environment—a review. *Annals of Animal Science*, 24(4), 1037-1049.
- Ashong, G. W., Ababio, B. A., & Kwaansa-Ansah, E. E. (2024). Evaluation of trace metals and quality of selected brands of vegetable cooking oils available on the Ghanaian market. *Journal of Trace Elements and Minerals*, 8, 100119.

- Ataei, A. H., Moheghi, M. M., and Fazel, Y. (2022). Effect of grower dietary energy level on feed intake and performance of modern broiler chickens. *Journal of Poultry Research*, 19(1), 1–6.
- Attia, Y. A., Al-Harhi, M. A., and Abo El-Maaty, H. M. (2015). The effects of dietary supplementation with phytase enzyme on laying performance, egg quality, bone mineralization and phosphorus excretion in laying hens. *Animal Physiology and Animal Nutrition*, 99(3), 620–628.
- Avedeweh, A. E. (2015). Effect of heating time on yield and quality of soya bean meal and oil (Doctoral dissertation).
- Axentii, M., and Codină, G. G. (2024). Exploring the Nutritional Potential and Functionality of Hemp and Rapeseed Proteins: A Review on Unveiling Anti-Nutritional Factors, Bioactive Compounds, and Functional Attributes. *Plants*, 13(9), 1195.
- Azman, M. A., and Çiftçi, M. (2020). Effects of dietary fat sources on liver and serum cholesterol levels of broilers. *Journal of Poultry Research*, 17(1), 25-32.
- Babatunde, O. O., Park, C. S., and Adeola, O. (2021). Nutritional potentials of atypical feed ingredients for broiler chickens and pigs. *Animals*, 11(5), 1196.
- Badu-Apraku, B., and Fakorede, M. A. B. (2017). Maize in Sub-Saharan Africa: importance and production constraints. In *Advances in genetic enhancement of early and extra-early maize for Sub-Saharan Africa* (pp. 3-10). *Springer, Cham*.
- Baéza E, Le Bihan-Duval E. Chicken lines divergent for low or high abdominal fat deposition: A relevant model to study the regulation of energy metabolism. *Animal*. 2013;7:965-973

- Baião NC, Lara L. 2005. Oil and fat in broiler nutrition. *Revista Brasil Ciên Avíc.* 7:129–141.
- Bankole, J. A. (2022). Growth and yield of sorghum cultivars as influenced by population density of components of soya bean (Master's thesis, Kwara State University (Nigeria)).
- Barbour, G. W., Farran, M. T., Usayran, N. N., Darwish, A. H., Uwayjan, M. G., & Ashkarian, V. M. (2006). Effect of soybean oil supplementation to low metabolizable energy diets on production parameters of broiler chickens. *Journal of applied poultry research*, 15(2), 190-197.
- Barbour, G. W., Farran, M. T., Usayran, N. N., Darwish, A. H., Uwayjan, M. G., and Ashkarian, V. M. (2006). Effect of soya bean oil supplementation to low metabolizable energy diets on production parameters of broiler chickens. *Journal of applied poultry research*, 15(2), 190-197.
- Barnes, S. (2010). The Biochemistry, chemistry, and Physiology of the isoflavones in soya beans and their food products. *Journal of Nutrition*, 1325S-1329S.
- Barros, T., Quaassdorff, M. A., Aguerre, M. J., Colmenero, J. O., Bertics, S. J., Crump, P. M., and Wattiaux, M. A. (2017). Effects of dietary crude protein concentration on late-lactation dairy cow performance and indicators of nitrogen utilization. *Journal of Dairy Science*, 100(7), 5434-5448.
- Batal, A. B., and Parsons, C. M. (2004). Utilization of various carbohydrate sources as energy by broiler chicks in the first week of life. *Poultry Science*, 83(3), 423–428. <https://doi.org/10.1093/ps/83.3.423>

- Batal, A., and Dale, N. (2016). Ingredient analysis table: 2016 edition. Feedstuffs Reference Issue.
- Becquet, P., Vazquez-Anon, M., Mercier, Y., Wedekind, K., Mahmood, T., Batonon-Alavo, D. I., and Yan, F. (2023). A systematic review of metabolism of methionine sources in animals: One parameter does not convey a comprehensive story. *Animal Nutrition*, 13, 31-49.
- Bello, A. U., and Olomu, J. M. (2023). Nutritional evaluation of full-fat soya bean meal for broilers: Effect on performance, nutrient digestibility, and carcass quality. *Tropical Animal Health and Production*, 55(1), 34. <https://doi.org/10.1007/s11250-023-03558-9>
- Bender, M. F., Schmidt, D. A., and Vogel, M. J. (2021). Effect of heat-treated unrefined soya beans on broiler performance and carcass characteristics. *Journal of Animal Science*, 99(10), 2017-2023.
- Bergeron, A. N. (2021). Evaluate the Effects of Fat Sources and Different Calcium Solubility on Broiler Performance, Nutrient Absorption, Intestinal pH, and Bone Development (Doctoral dissertation).
- Bingol, N. T., Dede, S., Karsli, M. A., Değer, Y., Kılınç, K. D., and Kiliçalp, S. (2016). Effects of the replacement of soya bean meal with pea as dietary protein source on the serum protein fractions of broilers. *Brazilian Journal of Poultry Science*, 18, 639-644.
- Bora, P. (2014). Anti-nutritional factors in foods and their effects. *Journal of Academia and Industrial Research*, 3(6), 285-290.

- Bougouin, A., Appuhamy, J. A. D. R. N., Kronberg, S. L., and Kebreab, E. (2015). Phytate degradation in the rumen and hindgut of dairy cows: A meta-analysis. *Journal of Dairy Science*, *98*(10), 7450–7462.
- Calder, P. C (2015). Functional roles of fatty acids and their effects on human health. *Journal of Parenteral and Enteral Nutrition*, *39*(1\_suppl), 18S-32S.
- Cao, M. H., and Adeola, O. (2016). Energy value of poultry byproduct meal and animal-vegetable oil blend for broiler chickens by the regression method. *Poultry Science*, *95*(2), 268-275.
- Carmona, J. M., Lopez-Bote, C. J., Daza, A., and Rey, A. I. (2019). Fat accumulation, fatty acids and melting point changes in broiler chick abdominal fat as affected by time of dietary fat feeding and slaughter age. *British poultry science*, *60*(3), 219-228.
- Castro, T., Martinez, D., Isabel, B., Cabezas, A., and Jimeno, V. (2019). Vegetable oils rich in polyunsaturated fatty acids supplementation of dairy cows' diets: Effects on productive and reproductive performance. *Animals*, *9*(5), 205.
- Chandran, A. S., Suri, S., and Choudhary, P. (2023). Sustainable plant protein: an up-to-date overview of sources, extraction techniques and utilization. *Sustainable Food Technology*, *1*(4), 466-483.
- Chatterjee, R. N., and Rajkumar, U. (2015). An overview of poultry production in India. *Indian Journal of Animal Health*, *54*(2), 89-108.
- Chaudhry, A. S., Pasha, T. N., and Iqbal, Z. (2015). Nutritional evaluation of full-fat soya beans in broiler diets. *Journal of Animal and Feed Sciences*, *24*(1), 15–21.
- Chaudhry, R., & Varacallo, M. (2018). *Biochemistry, glycolysis*.

- Chen, B., and Ellefson, W. C. (2024). Fat Analysis. In Nielsen's Food Analysis (pp. 273-286). Cham: Springer International Publishing.
- Chen, P., Shen, Z., Ming, L., Li, Y., Dan, W., Lou, G., and He, Y. (2018). Genetic basis of variation in rice seed storage protein (Albumin, Globulin, Prolamin, and Glutelin) content revealed by genome-wide association analysis. *Frontiers in plant science*, 9, 612.
- Choct, M. (2015). Feed non-starch polysaccharides for monogastric animals: classification and function. *Animal Production Science*, 55(12), 1360-1366.
- Choct, M. (2018). Managing gut health through nutrition. *British Poultry Science*, 59(4), 468-474.
- Choi, J., Kong, B., Bowker, B. C., Zhuang, H., & Kim, W. K. (2023). Nutritional strategies to improve meat quality and composition in the challenging conditions of broiler production: a review. *Animals*, 13(8), 1386.
- Chrystal, P. V., Moss, A. F., Khoddami, A., Naranjo, V. D., Selle, P. H., & Liu, S. Y. (2020). Effects of reduced crude protein levels, dietary electrolyte balance, and energy density on the performance of broiler chickens offered maize-based diets with evaluations of starch, protein, and amino acid metabolism. *Poultry Science*, 99(3), 1421-1431.
- Chwen, LT, Foo HL, Thanh NT, Choe D. (2013). Growth performance, plasma fatty acids, villous height and crypt depth of preweaning piglets fed with medium chain triacylglycerol. *Asian-Aus J Anim Sci*. 26:700–704.
- Classen, H. L. (2017). Diet energy and feed intake in chickens. *Animal Feed Science and Technology*, 233, 13-21.

- Couce, M. L., & Saenz de Pipaon, M. (2021). Bone mineralization and calcium phosphorus metabolism. *Nutrients*, 13(11), 3692.
- Cowieson, A. J., and Adeola, O. (2005). The effect of enzyme supplementation on energy utilization and performance of poultry. *World's Poultry Science Journal*, 61(1), 85–96.
- Cowieson, A. J., Ptak, A., Mackowiak, P., Sassek, M., and Świątkiewicz, S. (2017). The effect of exogenous enzymes on performance and nutrient utilization in broilers fed wheat-based diets. *Poultry Science*, 96(9), 3145–3153.
- Crespo, N., & Esteve-Garcia, E. (2001). Dietary fatty acid profile modifies abdominal fat deposition in broiler chickens. *Poultry science*, 80(1), 71-78.
- Crespo, N., and Enric Esteve-Garcia. "Dietary fatty acid profile modifies abdominal fat deposition in broiler chickens." *Poultry science* 80.1 (2001): 71-78.
- Das, K. C. (2019). Effect Of DDGS as a replacer of soya bean meal on sonali chicken (Doctoral Dissertation, Hajee Mohammad Danesh Science and Technology University, Dinajpur.).
- De Coca-Sinova, A., Valencia, D. G., Jiménez-Moreno, E., *et al.* (2020). Effect of feed form and enzyme supplementation on nutrient digestibility and digestive pH in broilers. *Animal Feed Science and Technology*, 263, 114478.
- De Groot, S., Kersjes, J., Natonek, V., Roefs, B., and Scavizzi, S. (2022). The growing competition between the bioenergy industry and the feed industry.
- Dei, H. K. (2011). Soya bean as a feed ingredient for livestock and poultry (pp. 215-226). London: Intech Open.
- Dei, H. K. (2017). Assessment of maize (*Zea mays*) as feed resource for poultry. *Poultry*

- Dela Cruz, J. F., Acda, S. P., Centeno, J. R., and Carandang, N. F. (2014). Effects of different corn hybrids on performance parameters, carcass yield and organoleptic characteristics of broilers. *Philippine Journal of Veterinary and Animal Sciences*, 38(1).
- Dlamini, N. M., Madumo, T. S., and Nhlengethwa, S. L. (2020). The effects of moringa seed cake inclusion on the growth performance of broilers. *Poultry Science Journal*, 99(8), 3567-3576.
- Eleroglu, H., Karaca, A. C., and Savaş, T. (2019). The effects of pelleted and mash feed on broiler performance. *Poultry Science Journal*, 98(12), 5102-5108.
- Eleroglu, H., Karaca, A. C., and Savaş, T. (2020). The effect of different soya bean meal types on the carcass characteristics and growth performance of broilers. *Animal Feed Science and Technology*, 272, 114714.
- El-Safty, S. A., Hassanein, K. M. A., and Abdel-Wareth, A. A. A. (2022). Influence of soya bean processing on performance, carcass traits, and biochemical profile in broilers. *Livestock Science*, 257, 104870. <https://doi.org/10.1016/j.livsci.2022.104870>
- El-Wardany, I., Ali, H. F., and Ahmed, M. (2023). Effects of dietary enzymes on intestinal pH, microbial population, and bone quality in broilers. *Animal Nutrition and Health*, 12(3), 115–123.
- Enyisi, I. S., Umoh, V. J., Whong, C. M. Z., Alabi, O., Abdullahi, I. O. (2014). Chemical and nutritional values of maize and maize products obtained from selected markets in Kaduna. *Journal of Pharmaceutical and Allied Sciences*. 2014;11(2):2106-2113.

- Erdaw, M. M., Bhuiyan, M. M., and Iji, P. A. (2016). Enhancing the nutritional value of soya beans for poultry through supplementation with new-generation feed enzymes. *World's Poultry Science Journal*, 72(2), 307-322.
- Erenstein, O., Jaleta, M., Sonder, K., Mottaleb, K., & Prasanna, B. M. (2022). Global maize production, consumption and trade: trends and R&D implications. *Food security*, 14(5), 1295-1319.
- Esonu, B. O., Opara, M. N., Okoli, I. C., and Udedibie, A. B. I. (2017). Evaluation of full-fat soya bean meal as an alternative protein source for broilers. *International Journal of Poultry Science*, 16(2), 45–51.
- Eswaran, S., Muir, J., and Chey, W. D. (2013). Fiber and functional gastrointestinal disorders. *Official Journal of the American College of Gastroenterology|ACG*, 108(5), 718-727.
- Fan, Y., Zhao, L., Ma, Q., Li, X., and Wang, X. (2008). Effects of replacing corn with soya bean oil and soya beans on growth performance and carcass characteristics of broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 21(8), 1165–1171. <https://doi.org/10.5713/ajas.2008.70616>
- Feizollahi, E., Mirmahdi, R. S., Zoghi, A., Zijlstra, R. T., Roopesh, M. S., and Vasanthan, T. (2021). Review of the beneficial and anti-nutritional qualities of phytic acid, and procedures for removing it from food products. *Food Research International*, 143, 110284.
- Fernando, H. S. S. (2017). Effects of pretreatments on separating the seed coat from the cotyledon of black bean (Master's thesis, North Dakota State University).

- Forbes, J. M. (2007). *Voluntary Food Intake and Diet Selection in Farm Animals*. CAB International.
- Fritts, C. A., and Waldroup, P. W. (2003). Effect of source and level of vitamin D on live performance and bone development in growing broilers. *Journal of Applied Poultry Research*, 12(1), 45–52. <https://doi.org/10.1093/japr/12.1.45>
- Fryer, M. S., Slaton, N. A., Roberts, T. L., and Ross, W. J. (2019). Validation of soil-test-based phosphorus and potassium fertilizer recommendations for irrigated soya bean. *Soil Science Society of America Journal*, 83(3), 825-837.
- Gebru, Y. A., Hyun-Ii, J., Young-Soo, K., Myung-Kon, K., and Kwang-Pyo, K. (2019). Variations in amino acid and protein profiles in white versus brown teff (*Eragrostis tef*) seeds, and effect of extraction methods on protein yields. *Foods*, 8(6), 202.
- Gharib-Naseri, K., Kheravii, S. K., and Swick, R. A. (2021). Gut development and function of broiler chickens under varying dietary and environmental conditions. *Animal Feed Science and Technology*, 275, 114883.
- Gheisari, A., Zarei, M., and Hajati, H. (2017). Dietary acidification and its influence on performance and bone mineralization in broiler chicks. *Journal of Applied Poultry Research*, 26(2), 191–198.
- Ghiselli, R. F., Barbosa, V. M., and Sartori, J. R. (2022). Effect of dietary fat sources on calcium absorption and bone quality in poultry. *Poultry Science*, 101(4), 101860.
- Gilbert, J. A., Bendsen, N. T., Tremblay, A., & Astrup, A. (2011). Effect of proteins from different sources on body composition. *Nutrition, Metabolism and Cardiovascular Diseases*, 21, B16-B31.

- Gonzalez-Esquerria, R., and Leeson, S. (2005). Effects of menhaden oil and flaxseed in broiler diets on sensory quality and lipid composition of poultry meat. *British Poultry Science*, 46(4), 461-467
- Grieshop, C. M., Fahey, G. C., and Merchen, N. R. (2003). Amino acid and fatty acid composition of soya bean meal prepared from genetically modified soya beans. *Journal of Animal Science*, 81(3), 729-736
- Gu, R., Chen, F., Liu, B., Wang, X., Liu, J., Li, P., and Yuan, L. (2015). Comprehensive phenotypic analysis and quantitative trait locus identification for grain mineral concentration, content, and yield in maize (*Zea mays* L.). *Theor. Appl. Genet.* 128(9): 1777-1789.
- Gupta, R. K., Gangoliya, S. S., and Singh, N. K. (2015). Reduction of phytic acid and enhancement of bioavailable micronutrients in food grains. *Journal of food science and technology*, 52(2), 676-684.
- Haile, M. (2020). Effects of feeding different levels of soybean meal on performance of broiler chickens (*doctoral dissertation, haramaya university*).
- Hailu, M., & Kelemu, K. (2014). Trends in soy bean trade in Ethiopia. *Research Journal of Agriculture and Environmental Management*, 3(9), 477-484.
- He, R., Zhu, D., Chen, X., Cao, Y., Chen, Y. and Wang, X. (2019). How the trade barrier changes environmental costs of agricultural production: An implication derived from China's demand for soya bean caused by the US-China trade war. *Journal of Cleaner Production*, 227, pp.578-588.

- He, W., Li, P., and Wu, G. (2021). Amino acid nutrition and metabolism in chickens. *Amino acids in nutrition and health: Amino acids in the nutrition of companion, zoo and farm animals*, 109-131.
- Heger, J., Wiltafsky, M., and Zelenka, J. (2016). Impact of different processing of full-fat soya beans on broiler performance.
- Hellin, J., Erenstein, O., Beuchelt, T., Camacho, C., and Flores, D. (2013). Maize stover use and sustainable crop production in mixed crop–livestock systems in Mexico. *Field Crops Research*, 153, 12-21.
- Hetland, H., Svihus, B., and Krogdahl, Å. (2004). Effects of oat hulls and wood shavings on digestion in broilers and layers fed diets based on whole or ground wheat. *British Poultry Science*, 45(4), 456–466.  
<https://doi.org/10.1080/00071660400001171>
- Heuzé V., Tran G., Lebas F., 2017. *Maize grain*. Feedipedia, a programme by INRAE, CIRAD, AFZ and FAO. <https://www.feedipedia.org/node/556> Last updated on September 7, 2017, 14:29
- Hossain, F., Sarika, K., Muthusamy, V., Zunjare, R. U., and Gupta, H. S. (2019). Quality protein maize for nutritional security. In *Quality breeding in field crops* (pp. 217-237). Springer, Cham.
- Hou, S. (2023). The influence of lipid types and lipid levels on the performance parameters, apparent metabolisable energy and ileal nutrient digestibility in day 1-21 broilers fed maize-soya bean based starter diets: a thesis presented in partial fulfilment of the requirements for the degree of Master of Science in Animal Science at Massey

University, Palmerston North, Manawatū, New Zealand (Doctoral dissertation, Massey University).

Hussein, E. O. S., Suliman, G. M., Alowaimer, A. N., Ahmed, S. H., Abd El-Hack, M. E., Taha, A. E., & Swelum, A. A. (2020). Growth, carcass characteristics, and meat quality of broilers fed a low-energy diet supplemented with a multienzyme preparation. *Poultry science*, 99(4), 1988-1994.

Hussein, E. O. S., Suliman, G. M., Alowaimer, A. N., Ahmed, S. H., Abd El-Hack, M. E., Taha, A. E., and Swelum, A. A. (2020). Growth, carcass characteristics, and meat quality of broilers fed a low-energy diet supplemented with a multienzyme preparation. *Poultry science*, 99(4), 1988-1994.

Ignjatovic-Micic, D., Vancetovic, J., Trbovic, D., Dumanovic, Z., Kostadinovic, M., & Bozinovic, S. (2015). Grain nutrient composition of maize (*Zea mays* L.) drought-tolerant populations. *Journal of agricultural and food chemistry*, 63(4), 1251-1260.

Jain, A., Rastogi, D., and Chanana, B. (2016). Corn-a vital crop for our economy. *Research Journal of Humanities and Social Sciences*, 7(3), 185-192.

Jiménez-Moreno, E., Gonzalez-Alvarado, J. M., de Coca-Sinova, A., Lazaro, R., and Mateos, G. G. (2019). Effects of type and level of fiber on growth performance and digestive traits of broilers. *Poultry Science*, 98(5), 2195–2205.

Kafle, A., Timilsina, A., Gautam, A., Adhikari, K., Bhattarai, A., and Aryal, N. (2022). Phytoremediation: Mechanisms, plant selection and enhancement by natural and synthetic agents. *Environmental Advances*, 8, 100203.

- Karunaratne, N. D., Wijesekara, S., and Jayasena, D. D. (2023). Full-fat soya bean as a protein and energy source in broiler diets: Performance and economic perspectives. *Journal of Applied Poultry Research*, 32(2), 100327. <https://doi.org/10.1016/j.japr.2023.100327>
- Katoch, R. (2022). Nutritional Quality Estimation of Forages. In *Nutritional Quality Management of Forages in the Himalayan Region* (pp. 225-278). Singapore: Springer Singapore.
- Kaul, J., Jain, K., & Olakh, D. (2019). An overview on role of yellow maize in food, feed and nutrition security. *International Journal of Current Microbiology and Applied Sciences*, 8(02), 3037-3048.
- Kaur, N., Agarwal, A., Sabharwal, M., and Jaiswal, N. (2021). Natural Food Toxins as Anti-Nutritional Factors in Plants and Their Reduction Strategies. *Food Chemistry: The Role of Additives, Preservatives and Adulteration*, 217-248.
- Kaur, R., Kaur, G., Vikal, Y., Gill, G. K., Sharma, S., Singh, J. and Chawla, J. S. (2020). Genetic enhancement of essential amino acids for nutritional enrichment of maize protein quality through marker assisted selection. *Physiology and Molecular Biology of Plants*, 26(11), 2243-2254.
- Kellems, R. O., and Church, D. C. (2010). *Livestock Feeds and Feeding* (6th ed.). Pearson Prentice Hall.
- Kerr, B. J., Dozier III, W. A., and Shurson, G. C. (2023). Nutrient composition and feeding value of soya bean meal. *Journal of Applied Poultry Research*, 32(3), 100321.
- Khan, S. H., Rehman, A., and Sultan, J. I. (2020). Economic aspects of using various processed soya bean meals in poultry diets. *Poultry Science Journal*, 8(2), 89–95.

- Khan, S., Qamar, H., and Ahmad, I. (2022). The synergistic effect of moringa and acidifiers on intestinal pH, nutrient digestibility, and bone strength in broilers. *Veterinary Sciences*, 9(8), 430.
- Khattak, F. M., Pasha, T. N., Hayat, Z., and Mahmud, A. (2018). Enzymes in poultry nutrition. *Journal of Animal Science and Technology*, 60(1), 1-10.
- Kim, E. J., Utterback, P. L., and Parsons, C. M. (2021). Nutritional evaluation of full-fat soya beans for poultry and swine. *Journal of Animal Science and Biotechnology*, 12, 40. <https://doi.org/10.1186/s40104-021-00556->
- Kogut, M. H., and Arsenault, R. J. (2016). Avian gut immunity and the impact of nutrition on the gut microbiome. *Annual Review of Animal Biosciences*, 4, 55–72.
- Kokoszyński, D., Bernacki, Z., and Stępińska, M. (2022). Carcass composition and meat quality of broiler chickens. *Animal Science Journal*, 93(1), e13501.
- Korver, D. R. (2005). Genetic and nutritional regulation of bone mineralization in broiler chickens. *Poultry Science*, 84(7), 1060–1068. <https://doi.org/10.1093/ps/84.7.1060>
- Kour, G. (2024). Effect of dietary inclusion of phytogetic feed additives on the performance of layer quail (*Doctoral dissertation, Sher-e-Kashmir University of Agricultural Sciences and Technology, Jammu (JandK)*).
- Krautwald-Junghanns, M. E., Sürrie, C., and Reese, C. (2022). Impact of feed processing on vitamin degradation in poultry feed. *Poultry Science*, 101(9), 101934.
- Kumar, A., Dash, G. K., Sahoo, S. K., Lal, M. K., Sahoo, U., Sah, R. P., ... and Lenka, S. K. (2023). Phytic acid: A reservoir of phosphorus in seeds plays a dynamic role in plant and animal metabolism. *Phytochemistry Reviews*, 22(5), 1281-1304.

- Kumar, D., and Jhariya, A. N. (2013). Nutritional, medicinal and economic importance of corn: A mini review. *Res J Pharm Sci.* 2319, 555X.
- Kumar, K., Dasgupta, C. N., and Das, D. (2014). Cell growth kinetics of *Chlorella sorokiniana* and nutritional values of its biomass. *Bioresource Technology*, 167, 358–366.
- Kumari, D., Chandrasekara, A., and Shahidi, F. (2019). Bioaccessibility and antioxidant activities of finger millet food phenolics. *Journal of Food Bioactives*, 6, 100– 109 <https://doi.org/10.31665/JFB.2019.6187>.
- Lammers, B. P., & Heinrichs, A. J. (2000). The response of altering the ratio of dietary protein to energy on growth, feed efficiency, and mammary development in rapidly growing prepubertal heifers. *Journal of Dairy Science*, 83(5), 977-983.
- Lee, H. S., Kim, B. G., and Stein, H. H. (2022). Nutrient composition and digestibility of soya bean meal in poultry diets. *Journal of Animal Science and Biotechnology*, 13(1)
- Leeson, S., and Summers, J. D. (2001). *Nutrition of the Chicken* (4th ed.). University Books.
- Levic, J., and Sredanovic, S. (2010). Heat treatments in animal feed processing.
- Liu, L., Ren, M., Ren, K., Jin, Y., and Yan, M. (2020). Heat stress impacts on broiler performance: a systematic review and meta-analysis. *Poultry Science*, 99(11), 6205–6211.
- Liu, Y., Hu, Y., and Zhang, X. (2021). Effects of citric acid and probiotics on growth performance and tibia quality in broilers. *Poultry Science Journal*, 59(4), 122–130.

- Mabelebele, M., Adeola, O., and Mlambo, V. (2021). Effects of full-fat soya beans on growth, nutrient utilization, and carcass yield in broilers. *Animals*, 11(4), 981. <https://doi.org/10.3390/ani11040981>
- Maharjan, P., Martinez, D. A., Weil, J., Suesuttajit, N., Umberson, C., Mullenix, G., and Coon, C. N. (2021). Physiological growth trend of current meat broilers and dietary protein and energy management approaches for sustainable broiler production. *Animal*, 15, 100284.
- Manceron, S., Ben-Ari, T., and Dumas, P. (2014). Feeding proteins to livestock: Global land use and food vs. feed competition. *OCL Oilseeds and fats crops and lipids*, 21(4), 10.
- Manu, F., Okai, D. B., Boateng, M., and Frimpong, Y. O. (2015). Nutrient composition, pest and microbial status and effects of maize on the growth performance, carcass characteristics and economic profiles of growing-finishing pigs. *African Journal of Food, Agriculture, Nutrition and Development*, 15(4), 10241-10254.
- Masey O'Neill, H. V., Smith, J. A., and Bedford, M. R. (2020). Multicarbohydrase enzymes for non-ruminants. *Asian-Australasian Journal of Animal Sciences*, 33(3), 387–399.
- Mashiloane, T., Mlambo, V., Mhlongo, G., Dibakoane, S. R., & Mnisi, C. M. (2025). Cowpeas versus soybeans: Can valorization bridge the nutritional gap for sustainable animal feeding systems in the Global South?. *Frontiers in Sustainable Food Systems*, 9, 1657018.

- Mawiya, W. (2016). Effect of genotype and plant population on growth, nitrogen fixation and yield of soya bean [*Glycine max* (L.) Merrill] in the Sudan Savanna Agro-Ecological Zone of Ghana (Doctoral dissertation).
- McLeod J. Nutritional factors influencing carcass fat in broilers—A review. *World's Poultry Science Journal*. 1982;38(3):194-200. DOI: 10.1079/WPS19820015
- Messina, M. (2010). Soya beans isoflavones exposure does not have feminizing effect on men. A critical examination of the clinical evidence fertility and sterility, *93*(7), 2095-2104.
- Messina, M. (2016) soy and health update: Evaluation of the clinical and epidemiologic literature. *Nutrient* 8(12), 754
- Meyer, M. M., and Bobeck, E. A. (2021). Growth performance of male broilers fed ExPress® soya bean meal and high-shear dry extruded corn. *Journal of Applied Poultry Research*, 30(4), 100191.
- Mishra, R., Tripathi, M. K., Sikarwar, R. S., Singh, Y., and Tripathi, N. (2024). Soya bean (*Glycine max* L. Merrill): A multipurpose legume shaping our world. *Plant Cell Biotechnol. Mol. Biol*, 25, 17-37.
- Moffett, J. R., Puthillathu, N., Vengilote, R., Jaworski, D. M., & Namboodiri, A. M. (2020). Acetate revisited: A key biomolecule at the nexus of metabolism, epigenetics and oncogenesis—Part 1: Acetyl-CoA, acetogenesis and acyl-CoA short-chain synthetases. *Frontiers in Physiology*, 11, 580167.
- Mohammed, A. A., Jacobs, J. A., and Coon, C. N. (2020). Gender differences in bone development and mineral retention in broilers. *International Journal of Poultry Science*, 19(7), 335–342.

- Murdia, L. K., Wadhvani, R., Wadhawan, N., Bajpai, P., and Shekhawat, S. (2016). Maize utilization in India: an overview. *American Journal of Food and Nutrition*, 4(6), 169-176.'
- Murshed, M. (2018). Mechanism of bone mineralization. *Cold Spring Harbor perspectives in medicine*, 8(12), a031229.
- Musigwa, S., Morgan, N., Swick, R., Cozannet, P., and Wu, S. B. (2021). Optimisation of dietary energy utilisation for poultry—a literature review. *World's Poultry Science Journal*, 77(1), 5-27.
- Muslyumova, D. M., Kurilkina, M. Y., Duskaev, G. K., and Zavyalov, O. A. (2021). A method for increasing the productivity of meat gobies thanks to the use of cavitated sunflower oil sludge in the diet. In *IOP Conference Series: Earth and Environmental Science* (Vol. 839, No. 2, p. 022055). IOP Publishing.
- Mwambo, F. M., Fürst, C., Nyarko, B. K., Borgemeister, C., and Martius, C. (2020). Maize production and environmental costs: Resource evaluation and strategic land use planning for food security in northern Ghana by means of coupled emergy and data envelopment analysis. *Land Use Policy*, 95, 104490.
- Nabizadeh, S., Shariatifar, N., Shokoohi, E., Shoeibi, S., Gavahian, M., Fakhri, Y., and Mousavi Khaneghah, A. (2018). Prevalence and probabilistic health risk assessment of aflatoxins B 1, B 2, G 1, and G 2 in Iranian edible oils. *Environmental science and pollution research*, 25, 35562-35570.
- Nalle, C. L., Ravindran, V., Ravindran, G., and Hendriks, W. H. (2015). The effect of soya bean protein fractions on amino acid digestibility in broiler chickens. *Poultry Science*, 94(6), 1347-1357.

- National Research Council, and Subcommittee on Poultry Nutrition. (1994). Nutrient requirements of poultry: 1994. National Academies Press.
- Ndolo, V. U., and Beta, T. (2013). Distribution of carotenoids in endosperm, germ, and aleurone fractions of cereal grain kernels. *Food chemistry*, 139(1-4), 663-671.
- Negari, B., Yusuf, Y., Hundie, D., Ameha, N., Kebede, K., Abrar, B., and Diba, D. (2023). A comparison of growth performance, feed intake, and feed efficiency of broiler chickens fed on commercial and farm-formulated diets. *East African Journal of Veterinary and Animal Sciences*, 7(1), 1–7.
- Nguyen, T. T., Le, P. H., and Choi, Y. H. (2021). Effects of processing on vitamin content in soya bean meal used for poultry feeding. *Poultry Science*, 100(3), 1121-1130.
- Noblet, J., Wu, S. B., and Choct, M. (2022). Methodologies for energy evaluation of pig and poultry feeds: a review. *Animal Nutrition*, 8, 185-203.
- NRC (1994). Nutrient Requirements of Poultry (9th rev. ed.). National Research Council. National Academies Press.
- Oduguwa, O. O., Fanimu, A. O., and Olayeni, T. B. (2014). Performance of broiler chickens fed diets containing heat-treated full-fat soya bean. *Archivos de Zootecnia*, 63(242), 301–310.
- Ogbuewu, I. P., & Mbajiorgu, C. A. (2023). Utilisation of cassava as energy and protein feed resource in broiler chicken and laying hen diets. *Tropical Animal Health and Production*, 55(3), 161.
- Ojebiyi, O. O., Akinlade, J. A., and Akinade, S. O. (2014). Economic viability of broilers fed diets containing full-fat soya beans. *Global Journal of Bioscience and Biotechnology*, 3(4), 391–394.

- Ojewola, G. S., and Uko, O. J. (2005). Comparative evaluation of nutrient and energy values of some oil seeds for chicks. *International Journal of Poultry Science*, 4(11), 860–863. <https://doi.org/10.3923/ijps.2005.860.863>
- Okonkwo, C. I., Ekenyem, B. U., and Nwokedi, C. I. (2023). Proximate Composition and Energy Value of Nigerian Soya bean-Based Poultry Feeds. *Nigerian Journal of Animal Production*, 50(2), 102–109.
- Olabode, A. D., & Okelola, O. E. (2021). Principles of Animal Production. *Agricultural Technology for Colleges*, 245.
- Olabode, A. D., & Okelola, O. E. (2021). Principles of Animal Production. *Agricultural Technology for Colleges*, 245.
- Oladokun, S. A., Akinsoyinu, A. O., and Aribido, S. O. (2017). Effect of feeding different protein sources on carcass characteristics of broiler chickens. *Nigerian Journal of Animal Production*, 44(1), 92–98.
- Oladokun, S., Jegede, A. V., and Akinleye, M. O. (2018). Effects of soya bean oil inclusion on growth performance and nutrient utilization in broiler chickens. *Tropical Animal Health and Production*, 50, 1515–1522.
- Olomu, J. M. (2011). *Monogastric Animal Nutrition: Principles and Practice* (2nd ed.). Jachem Publications.
- Olukosi, O. A., Adeola, O., and Cowieson, A. J. (2008). Energy utilization and growth performance of broilers fed diets containing soya bean oil or full-fat soya beans with or without multienzyme supplement. *Poultry Science*, 87(5), 1012–1020. <https://doi.org/10.3382/ps.2007-00311>

- Olukosi, O. A., and Adeola, O. (2017). Prediction of soya bean meal energy and amino acid digestibility in poultry. *Poultry Science*, 96(6), 1678–1686.
- Olukosi, O. A., and Cowieson, A. J. (2016). Interactive effects of age and dietary mineral content on nutrient digestibility and bone development. *British Poultry Science*, 57(6), 760–767.
- Olukosi, O. A., Cowieson, A. J., and Adeola, O. (2013). Energy utilization and growth performance of broilers fed diets supplemented with enzymes. *Poultry Science*, 92(9), 2351–2357. <https://doi.org/10.3382/ps.2013-03107>
- Oluwafemi, R. A., Afolayan, S. B., and Ojo, V. O. A. (2016). Nutrient composition and economic impact of various soya bean processing techniques in broiler production. *African Journal of Livestock Extension*, 14(2), 49–56.
- Onu, P. N., and Madubuiké, F. N. (2012). Performance and economic analysis of broilers fed enzyme supplemented full-fat soya bean diets. *International Journal of Poultry Science*, 11(5), 298–303.
- Ortiz-Martinez, M., Otero-Pappatheodorou, J. T., Serna-Saldívar, S. O., and García-Lara, S. (2017). Antioxidant activity and characterization of protein fractions and hydrolysates from normal and quality protein maize kernels. *Journal of Cereal Science*, 76, 85-91.
- Owusu, V. (2022). Effects of regular maize and different varieties of maize on the growth performance and carcass traits of broiler chickens (Doctoral dissertation, University of Education Winneba).

- Oyekunle, O. O., Adeola, O., and Omoniyi, I. S. (2021). Impact of xylanase supplementation on feed efficiency in broilers. *Poultry Science*, 100(5), 1564-1570.
- Panghal, A., Munezero, C., Sharma, P., & Chhikara, N. (2021). Cassava toxicity, detoxification and its food applications: a review. *Toxin Reviews*.
- Phillips, S. M. (2016). The impact of protein quality on the promotion of resistance exercise-induced changes in muscle mass. *Nutrition and metabolism*, 13(1), 64.
- Prabhu, D., Dawe, R. S., and Mponda, K. (2021). Pellagra a review exploring causes and mechanisms, including isoniazid-induced pellagra. *Photodermatology, Photoimmunology and Photomedicine*, 37(2), 99-104.
- Prandini, A., Sigolo, S., Morlacchini, M., Marocco, A., Pinto, M. Lo, Cattolica, U., Piacenza, C., Erbacee, C., and Cattolica, U. (2016). High-protein maize in diets for broilers High-protein maize in diets for broilers. *Italian Journal of Animal Science*, 10(4), 243–249.
- Prasanna, B. M. (2012). Diversity in global maize germplasm: characterization and utilization. *Journal of biosciences*, 37(5), 843-855.
- Purohit, P., Rawat, H., Verma, N., Mishra, S., Nautiyal, A., Bhatt, S., ... and Gupta, A. K. (2023). Analytical approach to assess anti-nutritional factors of grains and oilseeds: a comprehensive review. *Journal of Agriculture and Food Research*, 100877.
- Qamar, M. N., Nafees, H., & Feroz, S. A. (2025). Role of Protein Nutrition for Maintaining the Health of the Musculoskeletal System. In *Evaluating the Effectiveness of*

*Functional Ingredients in Sports Nutrition* (pp. 177-214). IGI Global Scientific Publishing.

Rahbar, B., Safdar, A. H. A., and Kor, N. M. (2014). Mechanisms through which fat supplementation could enhance reproduction in farm animal. *European Journal of Experimental Biology*, 4(1), 340-348.

Rahimi, G., Abolhasani, M., and Gholami, M. (2022). The effects of vitamin E on feed conversion and growth in broilers. *Poultry Science Journal*, 101(6), 1420-1427.

Raji, A. O., Adebiyi, O. A., and Ogunleke, F. O. (2021). Effect of processing methods on the utilization of full-fat soya beans by broilers. *International Journal of Poultry Science*, 20(2), 87-94.

Ramli, N., Jamaludin, A. A., & Ilham, Z. (2017). Mimosine toxicity in *Leucaena* biomass: A hurdle impeding maximum use for bio products and Bioenergy. *Int J Environ Sci Nat Resour*, 6, 1-5.

Rath, N. C., Huff, G. R., Huff, W. E., and Balog, J. M. (2000). Factors regulating bone maturity and strength in poultry. *Poultry Science*, 79(7), 1024–1032.  
<https://doi.org/10.1093/ps/79.7.1024>

Ravindran, V. (2016). Poultry feed availability and nutrition in developing countries. *FAO Animal Production and Health Paper*, No. 201. Rome: FAO.

Ravindran, V. (2023). Anti-nutritional factors in soya bean and their implications for poultry nutrition. *World's Poultry Science Journal*, 79(1), 19-34.  
<https://doi.org/10.1080/00439339.2023.2178847>

Ravindran, V., Abdollahi, M. R., and Bootwalla, S. M. (2016). Nutrient analysis, metabolizable energy, and amino acid digestibility of soya bean meals of different

origins for broilers. *Poultry Science*, 95(3), 613–620.  
<https://doi.org/10.3382/ps/pev378>

Ravindran, V., Abdollahi, M. R., and Bootwalla, S. M. (2017). Nutrient analysis, metabolisable energy and digestibility of phosphorus in soya bean meal for poultry. *Animal Feed Science and Technology*, 225, 1–10.

Ravindran, V., and Hendriks, W. H. (2020). Soya bean processing: Nutritional and anti-nutritional implications for poultry diets. *Animal Feed Science and Technology*, 267, 114585.

Ravindran, V., Tanchaoenrat, P., Zaefarian, F., and Ravindran, G. (2017). Effects of dietary energy and protein on performance and nutrient utilization in broilers. *Poultry Science*, 96(9), 2941-2950.

Ravindran, V., Tanchaoenrat, P., Zaefarian, F., and Ravindran, G. (2016). Fats in poultry nutrition: Digestive physiology and factors influencing their utilization. *Animal Feed Science and Technology*, 213, 1-21

Ravindran, V., Wu, Y. B., and Morel, P. C. H. (2006). Performance and nutrient utilization of broiler chickens as influenced by dietary energy and lysine levels. *Journal of Animal and Feed Sciences*, 15(2), 271–281.

Reza, M. T., Emerson, R., Uddin, M. H., Gresham, G., and Coronella, C. J. (2015). Ash reduction of corn stover by mild hydrothermal preprocessing. *Biomass Conversion and Biorefinery*, 5(1), 21-31.

Rojo, R., Afsharmanesh, M., and Walk, C. L. (2023). Phosphorus digestibility and the effect of phytate in soya bean meal-based diets fed to broilers. *Animal Nutrition*, 9, 59–66.

- Rostagno, H. S., Albino, L. F. T., Hannas, M. I., Gomes, P. C., de Oliveira, R. F., Lopes, D. C., ... and Barreto, S. L. T. (2017). *Brazilian Tables for Poultry and Swine: Composition of Feedstuffs and Nutritional Requirements* (4th ed.). UFV Press.
- Rouf Shah, T., Prasad, K., and Kumar, P. (2016). Maize—A potential source of human nutrition and health: A review. *Cogent Food and Agriculture*. 2(1):1166995.
- Sahito, H. A., Soomro, R. N., Memon, A., Abro, M. R., Ujjan, N. A., & Rahman, A. (2012). Effect of fat supplementation on the growth, body temperature and blood cholesterol level of broiler. *Glo. Adv. Res. J. Chem. and Mat. Sci*, 1(2), 023-034.
- Saki, A. A., Shariatmadari, F., and Haghbin Nazarpak, H. (2017). Effect of feed form and nutrient density on performance and bone characteristics in broiler chicks. *Journal of Animal Physiology and Animal Nutrition*, 101(5), e229–e239.
- Sakomura, N. K. (2004). Modeling energy utilization in broiler breeders, laying hens and broilers. *Brazilian Journal of Poultry Science*, 6, 1-11.
- Schedle, K. (2016). Sustainable pig and poultry nutrition by improvement of nutrient utilisation—A review. *Die Bodenkultur: Journal of Land Management, Food and Environment*, 67(1), 45-60.
- Schmidt, R. H., Marshall, M. R., and O’Keefe, S. F. (2020). Total fat. In *Analyzing Food for Nutrition Labeling and Hazardous Contaminants* (pp. 29-56). CRC Press.science, 1, 1-32.
- Selim, S., Abdel-Megeid, N. S., Khalifa, H. K., Fakiha, K. G., Majrashi, K. A., & Hussein, E. (2022). Efficacy of various feed additives on performance, nutrient digestibility, bone quality, blood constituents, and phosphorus absorption and utilization of broiler chickens fed low phosphorus diet. *Animals*, 12(14), 1742.

- Shaghaghian, S., McClements, D. J., Khalesi, M., Garcia-Vaquero, M., and Mirzapour-Kouhdasht, A. (2022). Digestibility and bioavailability of plant-based proteins intended for use in meat analogues: A review. *Trends in Food Science and Technology*, 129, 646-656.
- Shah, T. R., Prasad, K., and Kumar P. (2016). Maize- a potential source of human nutrition and health: A review. *Cogent Food and Agriculture*. 2:1-9.
- Sharma, V., Sharma, S., and Yadav, S. (2020). Influence of dietary pH modifiers on growth and tibia characteristics of broiler chickens. *Indian Journal of Poultry Science*, 55(2), 101–106.
- Sheikhhasan, B. S., Moravej, H., Shivazad, M., Ghaziani, F., Esteve-Garcia, E., and Kim, W. K. (2020). Prediction of the total and standardized ileal digestible amino acid contents from the chemical composition of soya bean meals of different origin in broilers. *Poultry science*, 99(10), 4947-4957.
- Shim, M. Y., *et al.* (2012). Genetic variation of bone strength, mineral density, and microstructure in meat-type chickens. *Poultry Science*, 91(9), 2015–2023.
- Sibbald, I. R. (1986). The T.M.E. system of feed evaluation: Methodology, feed composition data, and bibliography. *Agriculture Canada Technical Bulletin*, 1986-4E.
- Singh, A. K., Berrocso, J. F. D., and Kim, W. K. (2018). Soya bean hulls and their effect on intestinal morphology in broilers. *Animal Feed Science and Technology*, 240, 42–50.

- Singh, A. K., Singh, A., and Das, T. K. (2015). Effect of processing on nutritional quality of soya bean products. *Journal of Food Science and Technology*, 52(12), 7248-7258.
- Singh, P., Kesharwani, R. K., & Keservani, R. K. (2017). Protein, carbohydrates, and fats: Energy metabolism. In *Sustained Energy for Enhanced Human Functions and Activity* (pp. 103-115). Academic Press.
- Singh, P., Kesharwani, R. K., & Keservani, R. K. (2017). Protein, carbohydrates, and fats: Energy metabolism. In *Sustained Energy for Enhanced Human Functions and Activity* (pp. 103-115). Academic Press.
- Singh, P., Pandey, V. K., Sultan, Z., Singh, R., and Dar, A. H. (2023). Classification, benefits, and applications of various anti-nutritional factors present in edible crops. *Journal of Agriculture and Food Research*, 14, 100902
- Slavin, J. L. (2005). Dietary fibre and body weight. *Nutrition*, 21(3), 411-418.
- Slominski, B. A. (2018). Advances in the understanding of dietary fibre and its components in relation to the use of alternative feed ingredients in modern poultry and livestock production.
- Sogbesan, O. A., Ogunlade, I. O., and Idowu, A. A. (2022). The effect of pellet size and hardness on broiler growth and feed conversion. *Animal Feed Science and Technology*, 278, 114933.
- Soomro, R. N., Yao, J., Hu, R., Memon, A., Abbasi, I. H. R., Arain, M. A., ... and Soomro, A. A. (2016). Effects of dietary fat supplementation on hematology and growth trait in broiler chickens. *Advances in Animal and Veterinary Sciences*, 4, 518-526.

- Sundu, B., Kumar, A., and Dingle, J. (2006). Feeding value of co-products of palm oil. *World's Poultry Science Journal*, 62(3), 567–581.  
<https://doi.org/10.1079/WPS200612>
- Surai, P. F. (2016). Antioxidant systems in poultry biology: Superoxide dismutase. *Journal of Animal Research and Nutrition*, 1(1), 1–11.
- Suri, D. J., and Sherry, A. (2016). Effects of different processing methods on the micronutrient and phytochemical contents of maize: From A to Z. *Comprehensive Reviews in Food Science and Food Safety*. 15:1541-4337.
- Svihus, B. (2011). The gizzard: function, influence of diet structure and effects on nutrient availability. *World's Poultry Science Journal*, 67(2), 207–224.  
<https://doi.org/10.1017/S0043933911000249>
- Svihus, B. (2014). Function of the digestive system. *Journal of Applied Poultry Research*, 23(2), 306-314.
- Swiatkiewicz, S., Arczewska-Wlosek, A., and Jozefiak, D. (2017). Bone health and mineral utilization in broilers fed diets supplemented with organic minerals: a review. *Annals of Animal Science*, 17(2), 351–372.
- Szczuko, M., Kikut, J., Komorniak, N., Bilicki, J., Celewicz, Z., and Ziętek, M. (2020). The role of arachidonic and linoleic acid derivatives in pathological pregnancies and the human reproduction process. *International journal of molecular sciences*, 21(24), 9628.
- Tang, H. G., Zhao, Y. L., and Wu, D. Y. (2024). Nutrient composition of full-fat soya bean meal and its role in poultry nutrition. *Poultry Science Journal*, 103(2), 200–209.

- Tenza, T., Mhlongo, L. C., Ncobela, C. N., & Rani, Z. (2024). Village chickens for achieving sustainable development goals 1 and 2 in resource-poor communities: *A literature review. Agriculture, 14*(8), 1264.
- Thakur, A., Sharma, V., and Thakur, A. (2019). An overview of anti-nutritional factors in food. *Int. J. Chem. Stud, 7*(1), 2472-247.
- Thrane, M., Paulsen, P. V., Orcutt, M. W., and Krieger, T. M. (2017). Soy protein: Impacts, production, and applications. In *Sustainable protein sources* (pp. 23-45). Academic press.
- Tijani, L. A., Balogun, O. S., and Lawal, T. T. (2016). Economic analysis of broiler production using different feed types. *Journal of Agricultural Economics and Development, 5*(2), 18–24.
- Toomer, O. T., Oviedo, E. O., Ali, M., Patino, D., Joseph, M., Frinsko, M., and Mian, R. (2023). Current agronomic practices, harvest and post-harvest processing of soya beans (*Glycine max*)—A review. *Agronomy, 13*(2), 427.
- Toomer, O. T., Oviedo-Rondón, E. O., Ali, M., Joseph, M., Vu, T., Fallen, B., & Mian, R. (2024). Full-Fat Soybean Meals as an Alternative Poultry Feed Ingredient—Feed Processing Methods and Utilization—Review and Perspective. *Animals, 14*(16), 2366.
- Torok, V. A., Hughes, R. J., Mikkelsen, L. L., Perez-Maldonado, R., Balding, K., MacAlpine, R., Percy, N. J., and Ophel-Keller, K. (2011). Identification and characterization of potential performance-related gut microbiotas in broiler chickens across various feeding trials. *Applied and Environmental Microbiology, 77*(17), 5868–5878. <https://doi.org/10.1128/AEM.00165-11>

- Vagadia, B. H., Vanga, S. K., and Raghavan, V. (2017). Inactivation methods of soya bean trypsin inhibitor—A review. *Trends in Food Science and Technology*, 64, 115-125.
- Vorland, C. J., Stremke, E. R., Moorthi, R. N., & Hill Gallant, K. M. (2017). Effects of excessive dietary phosphorus intake on bone health. *Current osteoporosis reports*, 15(5), 473-482.
- Vorland, C. J., Stremke, E. R., Moorthi, R. N., and Hill Gallant, K. M. (2017). Effects of excessive dietary phosphorus intake on bone health. *Current osteoporosis reports*, 15(5), 473-482.
- Voss, G. B., Rodríguez-Alcalá, L. M., Valente, L. M. P., and Pintado, M. M. (2018). Impact of different thermal treatments and storage conditions on the stability of soya bean byproduct (okara). *Journal of Food Measurement and Characterization*, 12, 1981-1996.
- Walk, C. L., Ball, M. E. E., and Morgan, N. K. (2021). Phytate: Impact on nutrient utilization and methods for improving phosphorus availability in poultry. *Animal Feed Science and Technology*, 275, 114882.
- Wang, S., Zhang, B., Chen, T., Li, C., Fu, X., and Huang, Q. (2019). Chemical cross-linking controls in vitro fecal fermentation rate of high-amylose maize starches and regulates gut microbiota composition. *Journal of Agricultural and Food Chemistry*, 67(49), 13728-13736.
- Wang, Y., Xu, B., Wu, S., and Zhang, H. (2017). Influence of dietary fatty acid profile on meat quality and lipid metabolism in poultry. *Journal of Agricultural and Food Chemistry*, 65(21), 4530–4538.

- Wei, P., Liu, M., Chen, Y., Chen, D., and Li, D. (2013). Effects of soy isoflavones on bone mineral density in postmenopausal women: a meta-analysis of randomized controlled trials. *Bone*, 55(2), 317–322. <https://doi.org/10.1016/j.bone.2013.03.009>
- Whitehead, C. C. (2004). Overview of bone biology in the avian species. *Poultry Science*, 83(2), 193–199.
- Williams, K. T., Weigel, K. A., Coblenz, W. K., Esser, N. M., Schlessner, H., Hoffman, P. C., ... & Akins, M. S. (2022). Effect of diet energy level and genomic residual feed intake on bred Holstein dairy heifer growth and feed efficiency. *Journal of Dairy Science*, 105(3), 2201-2214.
- Windisch, W., Schedle, K., Plitzner, C., and Newbold, C. (2008). Use of phytogetic products as feed additives for swine and poultry. *Journal of Animal Science*, 86(14\_suppl), E140–E148.
- Wu, S. B., Swick, R. A., Noblet, J., Rodgers, N., Cadogan, D., and Choct, M. (2019). Net energy prediction and energy efficiency of feed for broiler chickens. *Poultry Science*, 98(3), 1222-1234.
- Yadav, S., and Jha, R. (2019). Strategies to modulate the intestinal microbiota and its role in nutrient utilization and gut health of poultry. *Frontiers in Veterinary Science*, 6, 46.
- Yahav, S., Straschnow, A., and Plavnik, I. (2005). The effect of age at exposure to high ambient temperature on the development of thermotolerance in broiler chickens. *Journal of Thermal Biology*, 30(6), 429–435.

- Yatharth, S., Nikita W., Sarla L., and Jain, S. K. (2022). Analysis of nutritional composition of popular maize varieties. *The Pharma Innovation Journal* SP-11(10): 238-241.
- Zeng, Q. F., Cherry, P., Doster, A., Murdoch, R., Adeola, O., and Persia, M. E. (2015). Effects of dietary energy and fat source on growth performance and carcass traits of broilers. *Journal of Applied Poultry Research*, 24(3), 373–383. <https://doi.org/10.3382/japr/pfv035>
- Zerehdaran, S., Aghaei, A., and Fathi, M. M. (2023). Genetic improvement of feed conversion ratio in broilers. *Poultry Genetics and Breeding*, 39(1), 42-49.
- Zhang, B., Haitao, H., Wang, C., and Li, F. (2011). Effects of full-fat soya bean and soya bean oil on broiler performance, carcass composition, and serum lipid levels. *Poultry Science*, 90(12), 2890–2896. <https://doi.org/10.3382/ps.2010-01265>
- Zhang, Y., Ji, W., Wu, Y., Han, H., Qin, J., and Wang, Y. (2016). Replacement of dietary fish meal by soya bean meal supplemented with crystalline methionine for Japanese seabass (*Lateolabrax japonicus*). *Aquaculture Research*, 47(1), 243-252.
- Zhang, Y., Liu, Y., and Yang, Z. (2022). Effects of methionine supplementation on broiler carcass yield and breast weight. *Poultry Science*, 101(6), 1420-1427.
- Zhao, F., Zhang, H. J., Wang, J., and Zhang, J. F. (2020). Effect of dietary soyabean oil on growth performance, nutrient digestibility, and carcass quality in broiler chickens. *Poultry science*, 99(5), 2567-2575.
- Zhao, J. P., Jiang, Y. B., Song, X. M., and Wang, K. H. (2009). Effects of different fat sources on growth performance, nutrient digestibility and abdominal fat deposition in broilers. *Animal Feed Science and Technology*, 153(3–4), 235–243. <https://doi.org/10.1016/j.anifeedsci.2009.07.003>

Zhu, X., Liu, J., and Yang, G. (2021). Effects of soya bean oligosaccharide, stachyose, and raffinose on growth performance and cecal microbiota in broiler chickens. *Animal Science Journal*, 92(1), e13668.

Zulkifli, I., Kamaruddin, S. R., and Soleimani, M. (2019). Heat stress and its effect on feed conversion efficiency in broilers. *Poultry Science Journal*, 98(4).

## APPENDICES

### Week 2

#### Appendix A1: Calculation of Livability d 0-14

$$\text{count } d14 \div \text{count } d0 \times 100$$

#### Appendix A2: Calculation of Body weight d 0-14

$$\text{Pen weight } d14 \div \text{count } d14$$

#### Appendix A3: Calculation of Gain d 0-14

$$(\text{Pen weight } d14 \div \text{count } d14) - (\text{Initial weight } d0 \div \text{count } d0)$$

#### Appendix A4: Calculation of Consumption d 0-14

$$\begin{aligned} &\text{Feed in } d0 - \text{Feed out } d9 + \text{Feed in } d9 - \text{Feed out } d13 + \text{Feed in } d13 \\ &\quad - \text{Feed out } d14 \end{aligned}$$

#### Appendix A5: Calculation of FCR d 0-14

$$\begin{aligned} &\text{Consumption } d0 - 14 \div (\text{Pen weight } d14 - \text{initial weight } d0 \\ &\quad + \text{dead bodyweight } d0 - 14) \end{aligned}$$

#### Appendix A6: Calculation of Feed intake d 0-14

$$\text{Gain } d14 \times \text{FCR } d14$$

### Week 4

#### Appendix B1: Calculation of Livability d 0- 28

$$\text{count } d28 \div \text{count } d0 \times 100$$

#### Appendix B2: Calculation of Body weight d 0- 28

$$\text{Pen weight } d28 \div \text{count } d28$$

#### Appendix B3: Calculation of Gain d 0- 28

$(Pen\ weight\ d28 \div count\ d28) - (Initial\ weight\ d0 \div count\ d0)$

Appendix B4: Calculation of Consumption d 14- 28

$Feed\ in\ d14 - Feed\ out\ d18 + Feed\ in\ d18 - Feed\ out\ d21 + Feed\ in\ d21$   
 $- Feed\ out\ d24 + Feed\ in\ d24 - Feed\ out\ d26 + Feed\ in\ d26$   
 $- Feed\ out\ d28$

Appendix B5: Calculation of FCR d 0-28

$(Consumption\ d0 - 14 + Consumption\ d14 - 28) \div (Pen\ weight\ d28$   
 $- initial\ weight\ d0 + dead\ bodyweight\ d0 - 14$   
 $+ dead\ bodyweight\ d14 - 28)$

Appendix B6: Calculation of Feed intake d 0-28

$Gain\ d28 \times FCR\ d28$

**Week 6**

Appendix C1: Calculation of Livability d 0-42

$count\ d42 \div count\ d0 \times 100$

Appendix C2: Calculation of Body weight d 0-42

$Pen\ weight\ d42 \div Count\ d42$

Appendix C3: Calculation of Gain d 0-42

$(Pen\ weight\ d42 \div Count\ d42) - (Initial\ weight\ d0 \div Count\ d0)$

Appendix C4: Calculation of Consumption d 28-42

$$\begin{aligned}
& \text{Feed in } d28 - \text{Feed out } 31 + \text{Feed in } d31 - \text{Feed out } d33 + \text{Feed in } d33 \\
& \quad - \text{Feed out } d36 + \text{Feed in } d36 - \text{Feed out } d39 + \text{Feed in } d39 \\
& \quad - \text{Feed out } d42
\end{aligned}$$

Appendix C5: Calculation of FCR d 0-42

$$\begin{aligned}
& (\text{Consumption } d0 - 14 + \text{Consumption } d14 - 28 + \text{Consumption } d28 - 42 \\
& \quad \div (\text{Pen weight } d42 - \text{initial weight } d0 + \text{dead bodyweight } d0 - 14 \\
& \quad + \text{dead bodyweight } d14 - 28 + \text{dead body weight } d28 - 42)
\end{aligned}$$

Appendix C6: Calculation of Feed intake d 0-42

$$\text{Gain } d42 \times \text{FCR } d42$$

**Week 8**

Appendix D1: Calculation of Livability d 0- 56

$$\text{count } d56 \div \text{count } d0 \times 100$$

Appendix D2: Calculation of Body weight d 0- 56

$$\text{Pen weight } d56 \div \text{Count } d56$$

Appendix D3: Calculation of Gain d 0- 56

$$(\text{Pen weight } d56 \div \text{Count } d56) - (\text{Initial weight } d0 \div \text{Count } d0)$$

Appendix D4: Calculation of Consumption d 42- 56

$$\begin{aligned}
& \text{Feed in } d42 - \text{Feed out } d45 + \text{Feed in } d45 - \text{Feed out } d49 + \text{Feed in } d49 \\
& \quad - \text{Feed out } d52 + \text{Feed in } d52 - \text{Feed out } d56
\end{aligned}$$

Appendix D5: Calculation of FCR d 0- 56

$$\begin{aligned}
& (\text{Consumption } d0 - 14 + \text{Consumption } d14 - 28 + \text{Consumption } d28 - 42 \\
& + \text{Consumption } d42 - 56 \div (\text{Pen weight } d56 - \text{initial weight } d0 \\
& + \text{dead bodyweight } d0 - 14 + \text{dead bodyweight } d14 - 28 \\
& + \text{dead body weight } d28 - 42 + \text{sample bird weight} \\
& + \text{dead body weight } d42 - 56
\end{aligned}$$

Appendix D6: Calculation of Feed intake d 42- 56

$$\text{Gain } d56 \times \text{FCR } d56$$